Recommended Nomenclature for Five Mammalian Carboxylesterase Gene Families: Human, Mouse and Rat Genes and Proteins

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Abstract

Mammalian carboxylesterase (*CES* or *Ces*) genes encode enzymes that participate in xenobiotic, drug and lipid metabolism in the body and are members of at least 5 gene families. Tandem duplications have added more genes for some families, particularly for mouse and rat genomes, which has caused confusion in naming rodent *Ces* genes. A new nomenclature system for human, mouse and rat carboxylesterase genes is described which identifies homolog gene families and allocates a unique name for each gene. The guidelines of human, mouse and rat gene nomenclature committees were followed and “*CES*” (human) or “*Ces*” (mouse and rat) root symbols were used followed by the family number (eg human *CES1*). Where multiple genes were identified for a family or where a clash occurred with an existing gene name, a letter was added (eg. human *CES4A*; mouse and rat *Ces1a*) which reflected gene relatedness among rodent species (eg. mouse and rat *Ces1a*). Pseudogenes were named by adding “*P*” and a number to the human gene name (eg. human *CES1P1*) or by using a new letter followed by *ps* for mouse and rat *Ces* pseudogenes (eg. *Ces2d-ps*). Gene transcript isoforms were named by adding the Genbank accession ID to the gene symbol (eg human *CES1_AB119995* or mouse *Ces1e_BC019208*). This nomenclature improves our understanding of human, mouse and rat *CES/Ces* gene families and facilitates research into the structure, function and evolution of these gene families. It also serves as a model for naming *CES* genes from other mammalian species.

INTRODUCTION

Five families of mammalian carboxylesterases (CES; E.C.3.1.1.1) have been described including CES1, the major liver enzyme (Munger et al., 1991; Shibata et al. 1993; Ghosh 2000; Holmes et al., 2009a); CES2, the major intestinal enzyme (Langmann et al. 1997; Schewer et al. 1997; Holmes et al., 2009a); CES3, expressed in brain, liver and colon (Sanghani et al. 2004; Holmes et al., 2010); CES5 (also called CES7 or cauxin), a major urinary protein of the domestic cat also present in human tissues (Miyazaki *et al.* 2003; 2006; Holmes et al., 2008a; Zhang et al., 2009); and CES6, a predicted CES-like enzyme in brain (Clark et al., 2003; Holmes et al., 2009a) (reviewed by Williams et al., 2010). These enzymes catalyse hydrolytic and transesterification reactions with xenobiotics, anticancer pro-drugs, and narcotics (Satoh & Hosokawa 1998; 2006; Satoh *et al.* 2002; Ohtsuka *et al.* 2003; Redinbo & Potter 2005), the conversion of lung alveolar surfactant (Ruppert et al. 2006) and several lipid metabolic reactions

(Tsujita & Okuda 1993; Becker et al. 1994; Ghosh 2000; Hosokawa et al. 2007; Diczfalussy et al. 2001), and may assist with the assembly of low density lipoprotein particles in liver (Wang et al. 2007).

Structures for human and animal *CES* genes have been reported, including rodent *CES1* and *CES2* ‘like’ genes (Ghosh et al. 1995; Dolinsky et al. 2001; Hosokawa et al. 2007) and human *CES1* and *CES2* genes (Becker et al. 1994; Langmann et al. 1997; Ghosh 2000; Marsh et al. 2004). Predicted gene structures have been also described for the human *CES3*, *CES5* and *CES6* genes, which are localized with *CES1* and *CES2* in two contiguous *CES* gene clusters on human chromosome 16 (Holmes et al. 2008a; 2009a,b; 2010). In addition, a *CES1*-like pseudogene (currently designated as *CES4*) is located with the *CES1-CES5* gene cluster (Yan et al., 1999). Mammalian *CES* genes usually contain 12 to 14 exons of DNA encoding CES enzyme sequences which may be shuffled during mRNA synthesis, generating several *CES* transcripts and enzymes encoded by each of the *CES* genes (see Thierry-Mieg and Thierry-Mieg, 2006). There are significant sequence similarities for the five *CES* families, especially for key regions previously identified for human liver CES1 (Bencharit et al. 2003; 2006; Fleming et al. 2005). Three-dimensional structural analyses of human CES1 have identified three major ligand binding sites, including the broad specificity active site, the ‘side door’ and ‘Z-site’, where substrates, fatty acids and cholesterol analogues respectively, are bound; and an active site ‘gate’, which may facilitate product release following catalysis (Bencharit et al. 2003; 2006; Fleming et al. 2005).

Because of the confusion associated with the current nomenclature for mammalian *CES* genes, particularly for mouse and rat CES genes where significant gene duplication events have generated a large number of *Ces1*-like and *Ces2*-like genes (see Berning et al., 1985; Ghosh et al., 1995; Satoh & Hosokawa, 1995; Dolinsky et al., 2001; Hosokawa et al., 2007), this paper proposes a new nomenclature system which enables easy identification of *CES* family members for this enzyme. The nomenclature follows the guidelines of the human, mouse and rat gene nomenclature committees and allocates a new name for each human (*CES*) or mouse and rat (*Ces*) gene. It also names and identifies the gene family origin for identified *CES* pseudogenes and provides a system for naming transcript isoforms derived from each of the *CES* genes. The nomenclature has the flexibility to accommodate new human, mouse and rat *CES* genes and will assist further research into the structure, function and evolution of these gene families as well as serving as a model for naming *CES* genes from other mammalian species.

GUIDING PRINCIPLES FOR THE NEW *CES* NOMENCLATURE

The new nomenclature system for human, mouse and rat *CES* genes and enzymes is based on the identification of homolog gene families and a subsequent allocation of a unique gene name for each of the genes observed from genome data bases or reported from previous studies. It follows the guidelines of the human, mouse and rat gene nomenclature committees and recommends the naming of homolog *CES* or *Ces* genes among species. The italicized root symbol “*CES*” for human and “*Ces*” for mouse and rat genes were used, followed by an number describing the gene family (examples include *CES1* for human *CES* family 1 or *Ces1* for mouse and rat *Ces* family 1 genes) (Tables 1-3). For mammalian genomes where multiple genes were identified or where a gene required a name which clashed with an existing name, a capital letter (for human genes) (eg. *CES4A*) or a lower case letter (for mouse and rat genes) (eg. *Ces1a*, *Ces1b* for multiple mouse *Ces1*-like genes) was added after the number. The letter used for multiple genes reflected the relatedness of the genes across species (eg. reflecting higher degrees of identity for mouse and rat *Ces1a* genes). When a human *CES* pseudogene was identified, a capital “P” and a number were added to the gene name (eg. *CES1P1*), whereas for mouse and rat *Ces* pseudogenes, a unique lower case letter was used followed by “-ps” (eg. *Ces2d-ps*). Transcript isoforms of human (*CES*), mouse and rat (*Ces*) gene transcripts were designated by following the gene name with the Genbank transcript ID, such as human *CES1_ABI19997* and *CES1_ABI187225* which differs from the current nomenclature used for human *CES1* isoforms (*CES1A1* and *CES1A2*, respectively) (see Table 1).

HUMAN *CES* GENES

Table 1 summarizes the locations and exonic structures for human *CES* genes based upon previous reports for human *CES1* and *CES2* (Becker et al. 1994; Langmann et al. 1997; Ghosh 2000; Marsh et al. 2004) and predictions for human *CES3* (Holmes et al., 2010), *CES4A* (Holmes et al., 2009a) and *CES5A* (Holmes et al., 2008a) (the February 2009 human reference sequence (GRCh37) was used in this study) (Rhead et al., 2010). Human *CES1P1* (a *CES1*-like pseudogene), *CES1* and *CES5A* were located in a cluster (cluster 1) on chromosome 16 while *CES2*, *CES3* and *CES4A* were in a separate cluster (cluster 2) on the same chromosome. Cluster 1 *CES* genes (*CES1* and *CES5A*) were transcribed on the negative strand, whereas cluster 2 genes (*CES2*, *CES3* and *CES4A*) were transcribed on the positive strand. Figure 1 summarizes the predicted exonic start sites for human *CES* genes, with *CES1* and *CES4A* containing 14 exons; *CES3* and *CES5A* 13 exons; and *CES2* with 12 exons. These exon start sites were in identical or similar positions to those reported for *CES1* (Ghosh, 2000; March et al., 2004). Figure 2 shows the comparative structures for human *CES* reference sequences and transcripts described

on the AceView website (<http://www.ncbi.nlm.nih.gov/IEB/Research/Acembly/>) (Thierry-Mieg & Thierry-Mieg, 2006). The *CES* gene and transcript sequences varied in size from 11kb for *CES2* to 79kb for *CES5A*, and exhibited distinct structures in each case. Moreover, several isoforms were generated *in vivo* for each of the human *CES* genes which have different structures, as a result of transcriptional events including truncation of the 5' ends, differential presence or absence of exons, alternative splicing or retention of introns or overlapping exons with different boundaries. In addition, the isoforms are differentially expressed in tissues of the body and may perform distinctive metabolic roles. *CES* isoforms were named by using the gene name followed by the Genbank ID for the specific transcript. Recent studies of human *CES1* have described at least two major isoform transcripts, designated as *CES1A1* (*AB119997*) and *CES1A2* (*AB119996*) (Tanimoto et al., 2007). These isoforms have been redesignated as *CES1_AB119997* and *CES1_AB119997*, respectively (see Table 1) and encode sequences which differ by only 4 amino acid residues within the N-terminal region (exon 1) (Tanimoto et al., 2007). Distinct 5'-untranslated consensus sequences for binding transcription factors were reported which suggested differences in transcriptional regulation and functional roles in contributing to CPT-11 chemosensitivity for these isoforms (Tanimoto et al., 2007; Hosokawa et al., 2008; Yoshimura et al., 2008). Fukami and coworkers (2008) have also examined human *CES* isoform structure and proposed that *CES1P1*, a *CES1*-like pseudogene on chromosome 16 (designated as *CES1A3*), was derived from the *CES1_AB119997* isoform.

HUMAN CES AMINO ACID SEQUENCES AND STRUCTURES

An alignment of the amino acid sequences for human CES-like protein subunits are shown in Figure 1, together with a description of several features for these enzymes. The sequences have been derived from previously reported sequences for *CES1* (Munger et al., 1991; Shibata et al., 1993); *CES2* (Langmann et al., 1997; Schewer et al., 1997); *CES3* (Sanghani et al., 2004); *CES4A* (previously *CES6* or *CES8*) (Holmes et al., 2009a); and *CES5A* (previously *CES7*) (Holmes et al., 2008a) (Table 1). Alignments of the human *CES* subunits showed between 39-46% sequence identities, which suggests that these are products of separate but related gene families, whereas sequence alignments of human *CES1* and *CES2* with mouse *CES1*-like and *CES2*-like subunits exhibited higher levels of sequence identities with the *CES* family homolog in each case (66-78% identities for human and mouse *CES1*-like subunits and 64-72% for human and mouse *CES2*-like subunits, respectively) (data not shown), suggesting that these are members of the same mammalian *CES* families, in each case. Similar results were observed for comparisons of human *CES3*, *CES4A* (previously *CES6* or *CES8*) and *CES5A* (previously

CES7) with the corresponding mouse CES homolog sequences, with 65%, 72% and 69% identities being observed, respectively. This supports the designation of these *CES* genes being members of the same family, in each case.

The amino acid sequences for the human CES subunits examined contained 567 (CES1), 559 (CES2), 571 (CES3), 561 (CES4A) and 575 (CES5A) residues (Figure 1). Previous studies on human CES1 have identified key residues which contribute to the catalytic, oligomeric, subcellular localization and regulatory functions for this enzyme (sequence numbers refer to human CES1). These included the catalytic triad for the active site (Ser221; Glu354; His468) (Cygler et al. 1993); disulfide bond forming residues (Cys87/Cys116 and Cys274/Cys285) (Lockridge et al. 1987); microsomal targeting sequences, including the hydrophobic N-terminus signal peptide (von Heijne 1983; Zhen et al. 1995; Potter et al. 1998) and the C-terminal endoplasmic reticulum (ER) retention sequence (His-Ile-Glu-Leu) (Robbi & Beaufay 1983); as well as ligand binding sites, including the 'Z-site' (Gly356), the 'side door' (Val424-Met425-Phe426) and the 'gate' (Phe550) residues (Bencharit et al. 2003; 2006; Fleming et al. 2005). Identical residues were observed for each of the human CES subunit families for the active site triad and disulfide bond forming residues although changes were observed for some key residues for CES1 subunits, including the 'side-door' and 'gate' of the active site, with family specific sequences or residues in each case; the 'Z-site' (Gly356 for human CES1) has been retained for human CES2 and CES5A sequences, but substituted for CES3 (Ser) and CES4A (Asn); the hydrophobic N-terminal sequence for human CES sequences has undergone major changes although this region retains a predicted signal peptide property; the human CES C-terminal tetrapeptide sequences have also changed, although CES2 (HTEL) and CES3 (QEDL) are similar in sequence with human CES1 (HIEL), which plays a role in the localization of human CES1 within endoplasmic reticulum membranes (Robbi & Beaufay, 1983).

Other key human CES1 sequences included two charge clamps which are responsible for subunit-subunit interaction, namely residues Lys78/Glu183 and Glu72/ which Arg186, contribute to the trimeric and hexameric structures for this enzyme (Bencharit et al. 2003; 2006; Fleming et al. 2005). Other human CES subunit sequences for these charge clamp sites included substitutions with neutral amino acids for the human CES2 and CES5A sequences, while the CES3 and CES4A sequences retained one potential clamp site (Figure 1). Pindel et al. (1997) and Holmes et al (2009b) have reported monomeric subunit structures for human and baboon CES2, which is consistent with the absence of charge clamps for this enzyme. This could have a major influence on the kinetics and biochemical roles for human CES isozymes since three dimensional studies have

indicated that ligand binding to the human CES1 ‘Z-site’ shifts the trimer-hexamer equilibrium towards the trimer facilitating substrate binding and enzyme catalysis (Redinbo & Potter 2005). The N-glycosylation site for human CES1 at Asn79-Ala80-Thr81 (Kroetz et al. 1993; Bencharit et al. 2003; 2006; Fleming et al. 2005) was not retained for any of the other human CES sequences, although potential N-glycosylation sites were observed at other positions, including CES2 (site 3), CES3 (site 2), CES4A (sites 4, 5 and 7) and CES5A (sites 6, 8 and 9) (Table 4). Given the reported role of the N-glycosylated carbohydrate group contributing to CES1 stability and maintaining catalytic efficiency (Kroetz et al. 1993), the N-glycosylation sites predicted for other human CES subunits may perform similar functions or indeed may serve new functions specific to a particular CES family.

Predicted secondary structures for human CES2 (Holmes et al., 2009b), CES3 (Holmes et al., 2010), CES4A (Holmes et al., 2009a) and CES5A (Holmes et al., 2008a) sequences were compared with those reported for human CES1 and similar α -helix β -sheet structures were observed for all of the CES subunits examined (Bencharit et al., 2003; 2006) (Figure 1). This was especially apparent near key residues or functional domains such as the α -helix within the N-terminal signal peptide; the β -sheet and α -helix structures near the active site Ser221 (human CES1) and ‘Z-site’ (Glu354/Gly356 respectively); the α -helices bordering the ‘side door’ site; and the α -helix containing the ‘gate’ residue (Phe550 for human CES1). The human CES5A sequence, however, contained a predicted helix at the hydrophobic C-terminus not observed for other CES subunits which may perform a family specific function. Predicted 3-D structures have been previously described for each of the human CES subunits (Holmes et al., 2008a; 2009a, b; 2010) which were similar to the human CES1 structure (Bencharit et al., 2003; 2006).

MOUSE *Ces* GENES AND ENZYMES

Table 2 summarizes the proposed names, locations and overall structures for the *Ces* genes observed for the mouse genome (July 2007 mouse [*Mus musculus*] genome data obtained from the Build 37 assembly by NCBI and the Mouse Genome Sequencing Consortium) (<http://www.ncbi.nlm.nih.gov> was used in this study). The italicized gene name, ‘*Ces*’, is consistent with other mouse gene nomenclature and is preferred to the ‘*CES*’ stem used for human genes. At least 20 mouse *Ces* genes are recognized on the Mouse Genome Database (<http://www.informatics.jax.org/>) (MGI) and further described in terms of their locations on mouse chromosome 8, the number of predicted exons for each gene, predicted strand for transcription, number of amino acid residues and subunit MWs for the encoded CES subunits, and identification symbols from MGI (eg.

MGI3648919 for *Ces1a*), NCBI (Reference Sequences were identified from the National Center for Biotechnology Information database) (<http://www.ncbi.nlm.nih.gov/>), Vega (the Vertebrate Genome Annotation (VEGA) database) (<http://vega.sanger.ac.uk/index.html>), UNIPROT (Universal Protein Resource) (<http://www.ebi.ac.uk/uniprot/>) and Ensembl (Genome Database) (<http://www.ensembl.org/>) database sources.

Eight *Ces1*-like genes are located in tandem within a 360 kilobase segment of mouse chromosome 8, with an average gene size of 28 kilobases. The names for these genes (*Ces1a*, *Ces1b*....*Ces1h*) are allocated in the same order as their locations on the mouse genome (Table 3). The *Ces1*-like gene cluster is also located near to the mouse *Ces5a* gene, which is comparable to the *CES1P1-CES1-CES5A* cluster observed for human chromosome 16. Each of these genes contained 13 or 14 exons predicted for transcription on the negative strand, and with encoded CES subunits exhibiting distinct but similar amino acid sequences (554-567 residues). The subunits were 63-85% identical with each other and with the human CES1 sequence, which is consistent with these being members of the mouse *Ces1* gene family. Mouse *Ces1*-like genes included several that have been previously investigated, including *Ces1c* (previously called *Es1*), encoding a major mouse plasma esterase with 554 amino acid residues and also exhibiting lung surfactant convertase activity (Genetta et al., 1988; Krishnasamy et al., 1998); *Ces1d* (previously *Ces3*), encoding a mouse liver enzyme with 565 residues exhibiting triacylglycerol hydrolase activity (Dolinsky et al., 2001); *Ces1e* (previously called *Es22* or *egasyn*) encoding a liver CES with 562 residues exhibiting β -glucuronidase-binding properties (Ovnic et al., 1991); and *Ces1g* (previously *Ces1*) encoding a liver CES which exhibits lipid metabolising activity and has 565 amino acid residues (Table 5) (Ellingham et al., 1998).

Eight *Ces2*-like genes were also observed in a second 286 kb gene cluster on mouse chromosome 8 with an average gene size of ~8 kilobases (Table 2). These genes were named according to their sequence of position on the mouse genome (*Ces2a*, *Ces2b*....*Ces2h*), and included a pseudogene designated as *Ces2d-ps*. Three of these mouse *Ces2*-like genes have been previously described including *Ces2c* (previously *Ces2*), which encodes an inducible liver acyl-carnitine hydrolase enzyme with 561 residues (Furihata et al., 2003); *Ces2e* (previously *Ces5*) encoding a liver and intestinal enzyme with 560 amino acid residues (The MGC Project Team, 2004); and *Ces2a* (previously *Ces6*), encoding a liver and colon enzyme with 558 residues (The MGC Project Team, 2004). The *Ces2*-like cluster was located alongside two *Ces3*-like mouse genes (*Ces3a* and *Ces3b*) and a *Ces4a* gene (Table 3), which is comparable with the *CES2-CES3-CES4A* gene cluster on human chromosome 16 (Table 1). The *Ces3a* gene (previously mouse *esterase 31*

or *Est31*) is expressed strongly in male mouse livers and encodes a 554 residue CES3-like subunit (Aida *et al.*, 1993), whereas the *Ces3b* gene (previously *Es31L* or *EG13909*) is also expressed in liver encoding a 568 residue subunit (The MGC Project Team, 2004). The *Ces4a* gene (previously called *EST8* or *Ces8*) encodes an enzyme predicted for secretion in epidermal cells with 563 amino acid residues showing 72% identity with human CES4A (The MGC Project Team, 2004).

RAT *Ces* GENES AND ENZYMES

Table 3 summarizes the proposed names, locations and structures for *Ces* genes observed for the rat genome (the November 2004 rat (*Rattus norvegicus*) genome assembly based on version 3.4 produced by the Baylor Human Genome Sequencing Center) (Gibbs *et al.*, 2004 was used in this study). Fifteen rat *Ces* genes were identified on the Rat Genome Database (RGD) (<http://rgd.mcw.edu/>) and further characterized by their locations on rat chromosomes 1 and 19, the number of predicted exons for each gene, the predicted strand for transcription, current gene symbols, the number of amino acid residues and subunit MWs for the encoded CES subunits, and the identification symbols from RGD (eg. RGD1583671 for *Ces1a*), NCBI Reference Sequences (<http://www.ncbi.nlm.nih.gov/>), Vega (<http://vega.sanger.ac.uk/index.html>), UNIPROT (<http://www.ebi.ac.uk/uniprot/>) and Ensembl (<http://www.ensembl.org/>) database sources.

Five *Ces1*-like genes were located in tandem within a 201 kilobase segment of rat chromosome 19, with an average gene size of 33 kilobases (Table 3). The names for these genes (*Ces1a*, *Ces1c*....*Ces1f*) were allocated according to their degree of identity with the corresponding mouse *Ces1*-like genes (Table 3). The genes were located in tandem in the same order as the mouse *Ces1*-like genes and were near the rat *Ces5a* gene. This is comparable to the *CES1P1-CES1A-CES5A* gene cluster observed for human chromosome 16. The rat *Ces1-like* genes contained 14 exons, and were predicted for transcription on the positive strand with encoded CES subunits exhibiting similar amino acid sequences (550-565 residues). The subunits were 65-73% identical with each other and with the human CES1 sequence, which is consistent with membership of the rat *Ces1* gene family. The encoded rat *Ces1-like* subunit sequences showed higher levels of identity with the corresponding mouse *Ces1-like* sequences (81-92% for rat and mouse CES1a, CES1c, CES1d, CES1e and CES1f amino acid sequences). At least 3 rat *Ces1-like* genes have been previously described, including *Ces1c* (previously called *Es1*) encoding a rat plasma esterase (Vanlith *et al.*, 1993; Sanghani *et al.*, 2002); *Ces1d* (previously *Ces3*) encoding a rat liver enzyme with 565 residues and exhibiting cholesteryl ester hydrolase

activity (Robbi et al., 1990; Ghosh et al., 1995); and *Ces1e* (previously called *ES-3* or *egasyn*), encoding a rat liver Ces with 561 residues and having β -glucuronidase-binding properties (Robbi & Beaufay, 1994).

Seven rat *Ces2*-like genes were observed on the rat genome which were localized on two chromosomes: chromosome 1 (*Ces2c* and *Ces2i*) and chromosome 19 in 3 locations: *Ces2a* and *Ces2e*; *Ces2j*; and *Ces2g* and *Ces2h* (Table 3). The genes were named according to the degree of sequence identity with the corresponding mouse *Ces2*-like genes. Rat *Ces2*-like genes have been previously investigated, including *Ces2c* (previously *Ces2*), encoding an inducible liver acyl-carnitine hydrolase enzyme containing 561 residues (Furihata et al., 2003); *Ces2e* (previously *Ces5*) encoding a liver and intestinal enzyme with 560 amino acid residues (The MGC Project Team, 2004); and *Ces2a* (previously *Ces6*), encoding a liver and colon enzyme with 558 residues (The MGC Project Team, 2004). The rat *Ces2*-like cluster was located alongside a *Ces3*-like gene (*Ces3a* and *Ces3b*) and a *Ces4a* gene (Table 3), which is comparable with the *CES2A-CES3A-CES4A* gene cluster on human chromosome 16 (Table 1).

FUNCTIONS OF MAMMALIAN CES FAMILIES

Mammalian CES families exhibit broad substrate specificities and specific roles for these enzymes have been difficult to establish because of the promiscuity of the CES active site towards a wide range of substrates and the existence of multiple forms with overlapping specificities (see Leinweber, 1987; Satoh & Hosokawa, 1998; 2006; Redinbo & Potter, 2005; Fleming et al., 2005; Imai, 2006). Table 4 summarizes current knowledge concerning substrates and functions reported for human, mouse and rat *CES* gene family members.

Studies on human CES1 have examined its role in the metabolism of various drugs, including narcotics such as heroin and cocaine (Pindel et al, 1997; Bencharit et al, 2003), warfare nerve agents (Hemmert et al., 2010), psychostimulants (Sun et al, 2004), analgesics (Takai et al., 1997) and chemotherapy drugs (Sanghani et al., 2004). Mammalian liver is predominantly responsible for drug clearance from the body with CES1 and CES2 (with CES1 > CES2) playing major roles, following absorption of drugs into the circulation (Pindel et al. 1997; Imai 2006). Mammalian intestine (with CES2 > CES1) plays a major role in first pass clearance of several drugs, predominantly via CES2 in the ileum and jejunum (Imai et al. 2003). CES1 and CES2 also have different roles in prodrug activation, as shown for the anti-cancer drug irinotecan (CPT-11) which is converted to its active form SN-38 predominantly by CES2 (Humerickhouse et al. 2000). Recent modeling studies have shown that the human CES2 active site cavity is lined

with negatively charged residues which may explain the preference of this enzyme for neutral substrates (Vistoli et al., 2010). The role for human CES3 has not been extensively studied although the enzyme is capable of activating prodrugs such as irinotecan (Sanghani et al., 2004). There are no reports concerning the metabolic role(s) for human CES4A and functional studies on mammalian CES5 function are limited to feline species, where the enzyme is secreted into cat urine and apparently regulates the production of a cat specific amino acid ‘felinine’, a putative pheromone precursor (Miyazaki et al. 2006).

EVOLUTION OF MAMMALIAN *CES* GENE FAMILIES

Recent comparative and evolutionary studies (Holmes et al., 2008b; Williams et al., 2010) have concluded that there at least five major mammalian *CES* gene families. In addition, the gene duplication events which generated the ancestral mammalian *CES1*, *CES2*, *CES3*, *CES4* and *CES5* genes have apparently predated the common ancestor for marsupial and eutherian mammals (Holmes et al., 2008b) which has been estimated at ~ 173-193 million years ago (Woodburne *et al*, 2003) and may coincide with the early diversification of tetrapods approximately 350-360 million years ago (Donoghue & Benton, 2007). The mammalian *CES* gene families are ancient in their genetic origins and were established prior to the appearance of mammals during evolution. Further *CES/Ces* gene duplication events have subsequently occurred during mammalian evolution however, especially for rodent species, for which the mouse and rat *Ces1*-like and *Ces2*-like genes have apparently undergone successive duplication events. At least three of these are likely to have occurred in the common ancestor for rat and mouse during rodent evolution since several homolog genes and proteins were recognized, including *Ces1c* (previously *Es1*), *Ces1d* (*Ces3*), *Ces1e* (*Es22*), *Ces2a* (*Ces6*), *Ces2c* (*Ces2*) and *Ces2e* (*Ces5*) (Tables 3 and 4). With the exception of the rat *Ces2*-like genes, which were located in multiple clusters on chromosomes 1 and 19, human, mouse and rat *CES* genes were localized within 2 clusters of genes on the same chromosome, namely *Ces1-Ces5A* (with multiple *Ces1*-like genes) and *Ces2-Ces3-Ces4A* (with multiple *Ces2*-like genes in mouse and rat). The presence of two *Ces3*-like genes in the mouse suggests that a further duplication event also took place in this species.

CONCLUSIONS

This paper has examined human, mouse and rat carboxylesterase genes and encoded subunits and has proposed a new nomenclature system identifying each of five gene families (designated as *CES1*, *CES2*...*CES5* for human genes or *Ces1*, *Ces2*...*Ces5* for mouse and rat genes) and allocating a unique gene name for each of the genes. The italicized root symbol “*CES*” for human and “*Ces*” for mouse and rat genes followed by a number for the family were used which is consistent with current practice. When multiple genes were identified for a gene family or where a gene required a name which clashed with an existing name, a capital letter (for human genes) (eg. *CES4A*) or a lower case letter (for mouse and rat genes) (eg. *Ces1a*, *Ces1b*) was added after the number. A human *CES* pseudogene was named using a capital “P” and a number (eg. *CES1P1*), whereas mouse and rat *Ces* pseudogenes were named with a unique lower case letter followed by “-ps” (eg. *Ces2d-ps*). This new nomenclature will also assist in naming multiple CES genes and proteins from other mammalian species. As an example, Holmes and co-workers (2009c) and Williams et al (2010) have reported multiple *CES1*-like genes on the horse genome which may be designated in accordance with the recommended nomenclature as *CES1A*, *CES1B*, *CES1C* etc in order of the tandem locations of these genes on chromosome 3. Transcript isoforms of *CES* gene transcripts were named by following the gene name with the Genbank ID for the specific transcript. This nomenclature will assist our understanding of the genetic relatedness and the CES family origins for individual human, mouse and rat CES genes and proteins and facilitate future research into the structure, function and evolution of these genes. It will also serve as a model for naming *CES* genes from other mammalian species.

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Figure Legends:

Figure 1: Amino Acid Sequence Alignments for Human CES1, CES2, CES3, CES4A and CES5A Subunits

See Table 1 for CES isoform sequences aligned; * shows identical residues for CES subunits; : similar alternate residues; . dissimilar alternate residues; signal peptide sequences for CES1 (1-17), CES2 (1-25), CES3 (1-27), CES4A (1-19) and CES5A (1-24) and C- termini (MTS) microsomal targeting sequences for CES1 (564-567), CES2 (556-569) and CES3 (568-571) are shown in **red**; active site (AS) triad residues (human CES1) Ser221, Glu354 and His468 are highlighted in **green**; 'Side door' (Val424-Met425-Phe426), 'Gate' (Phe550) and cholesterol binding residue (Z site) (Gly356) for human CES1 (Fleming et al, 2005) are highlighted in **khaki**; disulfide bond Cys residues for human CES1 (*) are shown in **blue**; charge clamp residues identified for human CES1 (Glu72...Arg186; Lys78...Glu183) (Fleming et al, 2005) are highlighted in **purple**; confirmed (CES1) (Asn79-Ala80-Thr81) [site 1] or predicted N-glycosylation sites for human CES2 (Asn111-Met112-Thr113) [site 3], CES3 (Asn105-Ser106-Ser107) [site 2], CES4A (Asn213-Val214-Thr215 [site 4]; Asn276-Ser-277-Thr278) [site 5]; and Asn388-Ile389-Thr390) [site 7] and CES5A (Asn363-Lys364-Ser365 [site 6]; Asn513-Leu514-Thr515 [site 8]; and Asn524-Met525-Ser526 [site 9]) are highlighted in **blue**; α -helix (human CES1 or predicted) and β -sheet (human CES1 or predicted) regions were highlighted in **yellow** and **grey**, respectively; α -helices and β -sheets are numbered according to the reported human CES1 3D structure (Fleming et al, 2005); **bold underlined** font shows known or predicted exon start sites; exon numbers refer to the human CES1 gene (see Langmann et al., 1997).

Figure 2: Gene Structures and Major Isoforms for Human CES1, CES2, CES3, CES4A and CES5A Genes.

Derived from AceView website <http://www.ncbi.nlm.nih.gov/IEB/Research/Acembly/> (Thierry-Mieg and Thierry-Mieg, 2006) Mature isoform variants (a) are shown with capped 5'- and 3'- ends for the predicted mRNA sequences; exons are in solid color; 5'- and 3'- untranslated regions of the genes are shown as open boxes; introns are shown as a line; the 5' → 3' transcription directions are shown; **a** refers to the major transcript isoform for each human CES gene. Note that each CES gene structure is drawn to a different scale and that the respective gene sizes are shown: CES1 [34.8kb]; CES2 [10.9kb]; CES3 [13.9kb]; CES4A [22.3kb]; and CES5A [79.3kb].

Table Legends

Table 1: Human *CES1*, *CES1P1*, *CES2*, *CES3*, *CES4A* and *CES5A* Genes and Subunits.

RefSeq, GenBank and UNIPROT IDs provide the sources for the gene and protein sequences; the relative gene expression level for human CES genes in comparison with the expression of an average human gene is given in [brackets] <http://www.ncbi.nlm.nih.gov/IEB/Research/Acembly/>; ²refers to GenBank ID number; ³no current AceView isoform name available; ps refers to a pseudogene (*CES1P1*); gene sizes are given as base pairs of nucleotides; +ve and -ve refer to the transcription strand direction; CES isoform sequences aligned in Figure 1 are shown in **bold**; ⁴the human CES2_BC032095 isoform transcript contains multiple transcription start sites with the shorter CES2 sequence (559 residues) previously reported (see Schwer et al., 1997; Pindel et al., 1997).

Table 2: Mouse *Ces* Genes and Subunits.

RefSeq, GenBank, UNIPROT, MGI, Vega and Ensembl IDs provide the sources for the gene and protein sequences; ps refers to a pseudogene (*Ces2d-ps*); [^]the relative gene expression level for mouse *Ces* genes in comparison with the expression of an average mouse gene is given in [brackets] <http://www.ncbi.nlm.nih.gov/IEB/Research/Acembly/>; gene sizes are given as base pairs of nucleotides; +ve and -ve refer to the transcription strand.

Table 3: Rat *Ces* Genes and Subunits.

RefSeq, GenBank, UNIPROT, RGD, Vega and Ensembl IDs provide the sources for the gene and protein sequences; gene sizes are given as base pairs of nucleotides; the relative gene expression level for rat *Ces* genes in comparison with the expression of an average rat gene is given in [brackets] <http://www.ncbi.nlm.nih.gov/IEB/Research/Acembly/>; +ve and -ve refer to the transcription strand direction.

Table 4: Functions and Substrates for Human *CES*, and Mouse and Rat *Ces* Genes and Enzymes

¹Pindel et al., 1997; ²Bencharit et al., 2003; ³Sato & Hosokawa, 2006; ⁴Sun et al., 2004; ⁵Takai et al., 1997; ⁶Humerickhouse et al., 2000; Xu et al., 2002; Ohtsuka et al., 2003; Morton et al., 2005; ⁷flurbiprofen derivatives serve as substrates; Imai, 2006; Taketani et al., 2007; Hosokawa, 2008; ⁸Diczfalussy et al., 2001; ⁹Hemmert et al., 2010; ¹⁰Mutch et al., 2007; ¹¹Becker et al., 1994; ¹²Barthel et al., 2008; ¹³Krishnasamy et al., 1998; Ruppert et al., 2006; ¹⁴Morton et al., 2005; ¹⁵Dolinsky et al., 2005; ¹⁶Ovnic et al., 1991; ¹⁷Ellingham et al., 1998; Koh et al., 2009; ¹⁸Furihata et al., 2003; ¹⁹Sanghani et al., 2002; ²⁰Ghosh et al., 1995; Okazaki et al., 2008; ²¹Robbi & Beaufay, 1994; ²²Masaki et al., 2007; ²³Miyazaki et al., 2006; ²⁴Ecroyd et al., 2006; Zhang et al., 2009 ²⁵Gilham et al., 2005; ²⁶Schreiber et al., 2009; ²⁷Lehner & Vance, 1999; ²⁸Okazaki et al., 2006; ²⁹Linke et al., 2005.

Figure 1

Figure 2

Table 1

[illegible]

Table 2

Mouse CES	Chromosome 8	Gene	Exons	Subunit	Amino	GenBank ID	MGI ID_YZ	Current MGI	Current	NCBI	Vega ID	Ensembl ID
Gene	Coordinates	Size	strand	MW	Acids			Symbol_YZ	Gene	Transcript		
(P roposed)		bps							Symbols			
Ces1a	95,544,116-95,572,091	27,979	14 -ve	61,744	563	BC089371	MGI:3648919	Gm4976	EG244595	NM_001013764	none	ENSMUSG00000071047
Ces1b	95,580,789-95,603,815	23,027	13 -ve	62,197	567	*NM_001081372	MGI:3779470	Gm5158	CesN	NM_001081372	none	ENSMUSG00000078964
Ces1c	95,622,914-95,655,182	32,268	13 -ve	61,172	554	BC028907	MGI:95420	Es1	Es1, Ces-N	NM_007954	ENSMUSG00000024453	ENSMUSG00000057400
Ces1d	95,690,157-95,721,618	31,462	14 -ve	61,788	565	BC019198	MGI:2148202	Ces3	Ces3	NM_053200	ENSMUSG00000024539	ENSMUSG00000056973
Ces1e	95,725,306-95,753,320	28,015	14 -ve	61,582	562	BC019208	MGI:95432	Es22	Es22	NM_133660	ENSMUSG00000024532	ENSMUSG00000061959
Ces1f	95,780,331-95,803,599	23,269	14 -ve	61,698	561	BC013479	MGI:234564	AU018778	CesML1, TGH-2	NM_144930	ENSMUSG00000024519	ENSMUSG00000031725
Ces1g	95,826,807-95,861,053	34,247	14 -ve	62,680	565	BC021150	MGI:88378	Ces1	Ces1	NM_021456	ENSMUSG00000024535	ENSMUSG00000057074
Ces1h	95,875,926-95,903,624	27,699	14 -ve	62,087	562	AK009689	MGI:75704	2310039D24Rik	AK009689	XM_134476	ENSMUSG00000033579	ENSMUSG00000074156
Ces2a	107,257,972-107,265,313	7,342	12 +ve	61,940	558	BC024491	MGI:2142491	Ces6	Ces6	NM_133960	OTTMUSG00000027410	ENSMUSG00000055730
Ces2b	107,355,572-107,362,353	6,782	12 +ve	61,927	556	BC015286	MGI:2448547	BC015286	BC015286	NM_198172	OTTMUSG00000027467	ENSMUSG00000050097
Ces2c	107,371,033-107,378,161	7,129	12 +ve	62,470	561	BC031170	MGI:2389505	Ces2	Ces2	NM_145603	OTTMUSG00000027466	ENSMUSG00000061825
Ces2d-ps	107,391,388-107,397,764	3,762	6 +ve			BC034182	MGI:3704319	Gm9756		XR_002069	none	ENSMUSG00000031884
Ces2e	107,450,221-107,457,611	7,391	12 +ve	62,735	560	BC055062	MGI:2443170	Ces5	Ces5	NM_172759	none	ENSMUSG00000031886
Ces2f	107,471,256-107,479,862	7,335	12 +ve	62,707	561	BC117742	MGI:1919153	2310038E17Rik		NM_001079865	none	ENSMUSG00000062826
Ces2g	107,485,688-107,492,328	6,771	10 +ve	52,731	478	BC027185	MGI:1919611	2210023G06Rik		NM_197999	none	ENSMUSG00000031877
Ces2h	107,524,753-107,544,307	19,554					MGI:3648740	Gm5744		XM_488149	none	none
Ces3a	107,572,572-107,582,000	21,512	13 +ve	61,510	554	AK138932	MGI:102773	Es31	Es31	NM_198672	none	ENSMUSG00000069922
Ces3b	107,607,670-107,617,468	9,799	14 +ve	63,007	568	BC019047	Gm4738	Es31L	Es31L	NM_144511	none	ENSMUSG00000062181
Ces4a	107,655,852-107,673,417	17,566	14 +ve	62,123	563	BC026374	BC026374	Ces8	Ces8	NM_146213	OTTMUSG00000027469	ENSMUSG00000060560
Ces5a	96,038,095-96,059,607	21,512	13 +ve	64,167	575	AB186393	MGI:1915185	Ces7	Ces7	NM_001003951	none	ENSMUSG00000058019

Table 3

[illegible]

Table 4

Mammal	CES (Ces)	Current Gene Symbol(s)	Substrates and Function (Hydrolysis or detoxification)	
	Gene			
Human	<i>CES1</i>	<i>CES1, hCE-1, CES1A1, HU1</i>	heroin, cocaine ¹⁻³ , methyl phenidate ⁴ , temocapril ⁵ , CPT-11 ⁶ , flurbiprofen ⁷	
		<i>CES1</i>	fatty acid ethyl ester synthase ⁸ , sarin ⁹ , ciclesonide ¹⁰ , cholesteryl ester hydrolase ¹¹ , triacylglycerol hydrolase ¹¹	
	<i>CES2</i>	<i>CES2, hCE-2, HU2</i>	procaine ⁵ , heroin, cocaine ¹⁻³ , temacapril ⁵ , CPT-11 ⁶ , flurbiprofen ⁷ , doxazolidine ¹²	
	<i>CES3</i>	<i>CES3</i>	CPT-11 ⁶	
Mouse	<i>Ces1c</i>	<i>Es1, Ces-N</i>	lung surfactant convertase ¹³ , CPT-11 ¹⁴	
	<i>Ces1d</i>	<i>Ces3</i>	triacylglycerol hydrolase ¹⁵	
	<i>Ces1e</i>	<i>Es22, egasyn</i>	β -glucuronidase binding in the liver endoplasmic reticulum ¹⁶ , retinyl ester hydrolase ²⁶	
	<i>Ces1f</i>	<i>CesML1, TGH-2</i>	triacylglycerol hydrolase ²⁷ , monoacylglycerol hydrolase ²⁷ , cholesteryl ester hydrolase ²⁷ , phospholipase ²⁷	
	<i>Ces1g</i>	<i>Ces1</i>	lipid metabolism ¹⁷	
	<i>Ces2c</i>	<i>Ces2</i>	inducible liver acylcarnitine hydrolase ¹⁸	
Rat	<i>Ces1c</i>	<i>Es1</i>	retinyl palmitate ¹⁹	
	<i>Ces1d</i>	<i>Ces3</i>	cholesterol ester hydrolase ²⁰ , triacylglycerol hydrolase ²⁷ , retinyl ester hydrolase ²⁸	
	<i>Ces1e</i>	<i>ES-3</i>	β -glucuronidase binding in the liver endoplasmic reticulum ²¹	
	<i>Ces2a</i>	<i>Ces6</i>	intestinal first pass metabolism ²²	
	<i>Ces2c</i>	<i>Ces2</i>	inducible liver acylcarnitine hydrolase ¹⁸ , intestinal first pass metabolism ²²	
	<i>Ces2e</i>	<i>Ces5</i>	intestinal first pass metabolism ²²	
Cat	<i>CES5A</i>	<i>CES7, cauxin</i>	3-methylbutanol-cysteinylglycine hydrolysis in urine releasing pheromone ²³	
Rat, sheep	<i>CES5A</i>	<i>CES7, cauxin</i>	lipid transfer reactions in epididymis ²⁴	