

# **Extraction and HPLC Characterisation of Chlorogenic Acid from Tobacco Residuals**

**(running title: Extraction and HPLC Characterisation of Chlorogenic Acid)**

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**Abstract:** Chlorogenic acid is a highly valuable natural polyphenol compound used in medicine and industries. Its current commercial sources are from plant extracts of *Lonicera japonica* Thunb and *Eucommia ulmoides* Oliver. These sources are limited and expensive. On the other hand, tobacco residuals contain chlorogenic acid and other natural polyphenol compounds. Large quantities of tobacco residuals are produced each year as waste materials from tobacco manufacturing, potentially providing an alternative commercial source of chlorogenic acid and other valuable compounds. In this paper, microwave and ultrasound extractions of chlorogenic acid with mixed solvent were studied. Total polyphenol

concentrations in extract solutions obtained with different extraction methods were analyzed with the method of ferrous tartrate and UV-Vis spectrophotometry and compared. The extraction solutions were also characterised for polyphenol compositions with the method of HPLC. Experimental results indicated that high extract concentrations of chlorogenic acid were obtained with a mixed solvent of acetone and water (1:2 v/v). A total polyphenol concentration of up to 4.87 mg/ml and a chlorogenic acid concentration of up to 2.12 mg/ml were achieved. The application of microwave and ultrasound significantly increased the extract concentrations. The extraction time needed was also much reduced. HPLC analysis indicated that acetone water mixed solvent extraction achieved much higher relative concentrations of chlorogenic acid to other compounds in the extract solutions. These results indicated that fast and effective extraction of chlorogenic acid from tobacco residuals were achieved.

**Keywords:** Tobacco residues, Polyphenol, Chlorogenic acid, Ferrous tartrate method, Solvent extraction, HPLC

## INTRODUCTION

Chlorogenic acid and many other polyphenol compounds are extensively used in medicine and industries such as in consumer chemicals and food industries (1). Chlorogenic acid is used as various additives in beverage, cosmetics, tea products and foods as well as medical substances (2-3). Chlorogenic acid has antibacterial and antiviral properties, and it is a natural antioxidant and anticancer agent (4). It is also a promising precursor compound for the development of medicine that can resist AIDS virus HIV (5). The current commercial sources of chlorogenic acids are from plant extracts of plants such as *Lonicera japonica Thunb* and

*Eucommia ulmoides* Oliver etc. (6-9). These sources are generally limited and therefore expensive.

Tobacco leaves contain chlorogenic acid and many other natural polyphenol compounds (10). Analytical data also indicated that the concentration of chlorogenic acid and rutin is the highest (75%-95%) among the polyphenol compounds in tobacco leaves (11-12). Therefore, this potentially provides an alternative commercial source for chlorogenic acid and other valuable polyphenol compounds (13). In addition, the tobacco industry produces large quantities of tobacco residues as waste materials, such as low grade tobacco leaves and tobacco powders. The extraction of solansol from tobacco leaves also produces large quantities of residues, which still contain the polyphenol compounds. The utilisation of these waste materials for the extraction of valuable compounds such as polyphenols and nicotine has the potential of significant environmental and economic benefits (14-15).

The objectives of this paper were two fold. The first was to study the extraction of chlorogenic acid and other polyphenol compounds from tobacco residues with a number of methods. The effectiveness of mixed solvent extraction with acetone and water and the use of ultrasound and microwave as extraction aids were investigated. The total concentrations of polyphenols in the extract solutions were analysed with the method of ferrous tartrate and UV-Vis spectrophotometry. The second objective was to characterise the polyphenol compositions of the extract solutions with the method of HPLC (16) and to study the effects of the extraction methods on the polyphenol compositions.

## **MATERIALS AND METHODS**

### **Tobacco Residues and Reagents**

The dry tobacco residues used in the experiments were obtained from Bangbu Cigarette Plant, Anhui, P.R. China. The tobacco leaves had been oven dried, sliced, re-moisturised and flavoured, and re-dried during the tobacco manufacturing processes. The residues were crushed into powders (2-3mm diameter) for use in the extraction experiments. Acetone, ethanol and  $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$ ,  $\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$ ,  $\text{KH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$  (analytical grade) were purchased from Nanjing Chemical Company. Rochelle salt (sodium potassium tartrate,  $\text{KNaC}_4\text{H}_4\text{O}_6 \cdot 4\text{H}_2\text{O}$ ; analytical grade) were purchased from Shenyang Chemical Reagent No.2 Plant. Chlorogenic acid standard was obtained from China Biological Products Assay Institute. Phosphoric acid and acetonitrile (chromatography grade) were purchased from Sigma Co.

A standard chlorogenic solution of 2 mg/ml was prepared by dissolving weighed amounts in ethanol. A ferrous tartrate solution was prepared by dissolving 0.1 g  $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$  and 0.5 g  $\text{KNaC}_4\text{H}_4\text{O}_6 \cdot 4\text{H}_2\text{O}$  in 100 ml distilled water. A phosphate buffer solution of pH 7.5 was prepared by adding 85 ml of a solution A (2.969 g  $\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$  dissolved in 250 ml distilled water) to 15 ml of a solution B (2.2695 g  $\text{KH}_2\text{PO}_4 \cdot 2\text{H}_2\text{O}$  dissolved in 250 ml of distilled water).

### **Analytical Instruments and Apparatus**

The following analytical instruments and apparatus were used in the experiments: HPLC system with UV detector and C18 reversed phase column (Model HP1100, Hewlett-Packard), UV-Vis spectrophotometer (Model 722, Shanghai No.3 Analytical Instrument Plant),

Incubator (Model PYX-DHS-50X65-S, Shanghai Yuejing Medical Appliance Plant), constant temperature water bath (Model HH-4, Changzhou Guohua Appliance Company), Microwave oven (Model WD900SL23-2,900 Watt), and Ultrasound Cleaner (Model KQ-100DB, 100Watt, Kunshan Ultrasound Instrument Company).

### 2.3 Analysis of Phenol Concentrations in Extract Solutions

The total polyphenol concentrations in the extract solutions were determined by the method of ferrous tartrate and UV-Vis spectrophotometry (16, 17). Chlorogenic acid standard solutions were used as the analysis standard. As the extract solutions contained chlorogenic acid and other polyphenol compounds, the measured concentrations can be regarded as the chlorogenic acid equivalent concentrations for the total concentrations of all the polyphenols in the extraction solutions. This equivalent concentration was referred as the total polyphenol concentration in the paper.

A calibration curved was prepared for the spectrophotometer analysis. The standard solutions for the calibration curve were prepared by diluting the 2 mg/ml chlorogenic acid standard solution. To each 25 ml test tube, 0, 0.25, 0.5, 0.75, 1.0, 1.25, and 1.5 ml chlorogenic acid standard solutions, respectively, were added. Distilled water was added to make up the liquid volume in each test tube to 1.5 ml. Then, 5 ml of ferrous tartrate solution and 10 ml of phosphate buffer solution were added to each test tube, and absorbance values at 540 nm were obtained to plot the calibration curve. Extract solution samples of 1.5 ml each were analysed with the same procedure.

### **Analysis of Polyphenol Composition in Extract Solutions**

The polyphenol compositions of the extract solutions were analysed with the method of HPLC. The HPLC conditions and procedures were as follows: MetaChem Polaris 5 $\mu$  C18-A silica gel column (5.0  $\mu$ m, 4.6 mm id  $\times$  250 mm), column temperature: 30 °C, mobile phase: 4:1 mixture solution of phosphate buffer solution (0.4%) ethylcyanide, flow rate: 0.7ml/min, and detection wavelength: 327nm. All samples solutions were filtered with Cellulose acetate filter paper (pore size 0.8  $\mu$ m) before HPLC analysis.

### **Extraction Experiments**

A number of methods were used for the extraction of chlorogenic acid and polyphenols from the tobacco residues. The extraction experiments were carried out as follows. The ratios of tobacco residue to solvent volume were fixed at 10 g to 90 ml in all extraction experiments. At the end of each extraction, the solutions were separated by filtration using filter paper and used for analyses. The extraction conditions used in the experiments were determined from a preliminary optimisation process and the extraction times for each of the methods were selected based on the estimation that most of the extractable polyphenols were extracted into the liquid phase and equilibrium conditions were achieved.

#### **Ethanol Extraction**

A sample of 10 g of tobacco residue powder was added into 90 ml of 95% ethanol in a flask with a reflux tube. The flask was then put in a hot water bath at 78 °C for 2 hr.

#### **Acetone Water Mixed Solvent Extraction**

A sample of 10 g of tobacco residue powder was added into 90 ml of acetone and water mixed solvent (1:2 v/v) in a beaker. The extraction was carried out for 2 days under static conditions.

### **Acetone Water Mixed Solvent Extraction with Ultrasound**

A sample of 10 g of tobacco residue powder was added into 90 ml of acetone and water mixed solvent (1:2 v/v) in a beaker. The extraction in the contents of the beaker was then carried out with ultrasound treatment (100 W) for a period of 15 min treatment.

### **Acetone Water Mixed Solvent Extraction with Microwave**

A sample of 10 g of tobacco residue powder was added into 90 ml of acetone and water mixed solvent (1:2 v/v) in a beaker. The extraction in the contents of the beaker was then carried out with microwave treatment (900 W) for a period of 90 sec. The temperature of the extract solution was increased because of the application of the microwave. However, due to the short duration of the extraction processes, there was no noticeable loss of the solvents.

## **RESULTS AND DISCUSSION**

### **Polyphenol Concentrations in Extract Solutions**

The effectiveness of a number of methods for the extraction of chlorogenic acid and other polyphenol compounds from tobacco residues was studied first. The total effective concentrations of polyphenol in the extraction solutions relative to standard chlorogenic acid as determined with the method of ferrous tartrate were obtained. These results were compared as in Table 1. The data given in Table 1 were average values from triplets and the standard deviation was in the order of 0.09 mg/ml.

It can be seen from Table 1 that all the extraction methods could effectively extract the polyphenol compounds from tobacco residues. However, there were large differences in extraction times that were needed. There were also some differences in the concentrations of

total polyphenol compounds in the extraction solutions. Ethanol is the most commonly used solvent in commercial extraction processes of chlorogenic acid products (5). Results in Table 1 showed that a concentration of 3.36 mg/ml was obtained with ethanol extraction for 2 hr at 78 °C. This was slightly less than 3.55 mg/ml obtained from acetone water (1:2 v/v) mixed solvent extraction at room temperature for 24 hr. In contrast, the applications of ultrasound and microwave not only dramatically reduced the extraction time needed, but also produced significantly higher concentrations of polyphenol compounds of 4.42 and 4.87 mg/ml, respectively, in the extraction solutions, indicating improvements in the recovery rates. It appears that ultrasound and microwave effectively disrupted the cellular organisational structures of tobacco leaves such that the polyphenol compounds were rapidly released into the solvent solutions. In addition, the improved extraction with ultrasound and microwave is also due to the improved mass transfer processes by such factors as cavitation effect, mechanic agitation effect and heat effect (19). Therefore, both ultrasound and microwave were demonstrated to be excellent aids for solvent extraction of compounds from tobacco residues, simultaneously with much reduced extraction time and significantly higher recovery rates.

### **Polyphenols Compositions in Extract Solutions**

Although the level of chlorogenic acid is relatively higher among the polyphenol compounds in tobacco leaves, the extraction solutions contain mixtures of polyphenol compounds. The actual compositions of the polyphenol compounds, as well as the effects of extraction on the resultant compositions are important and need to be characterised. In this paper, the characterisations of extraction solutions were achieved through the use of HPLC chromatograms, which were compared to the chromatogram of the standard solution of chlorogenic acid.

The chromatogram of the standard solution (2 mg/ml) is shown in Fig.1, in which the retention time of chlorogenic acid was at 5.6 min. There were no other peaks observed in the chromatogram, as standard chlorogenic acid solution was used.

The chromatogram of the extraction solution from ethanol extraction is shown in Fig.2. In Fig.2, it can be observed that in addition to the chlorogenic acid peak at 5.6 min, there are at least another 3 significant peaks before, and 2 after, the chlorogenic acid peak at retentions of 3.8, 4.2, 4.9, 8.9 and 9.9 min, indicating the presence of other substances such as other polyphenol compounds. The chlorogenic acid concentration in the extraction solution can be calculated from its peak area to be 0.54 mg/ml, which represents 26.6% of the total concentrations as calculated from the total peak areas in Fig.2. When compared to the total polyphenol concentration as determined by the ferrous tartrate method (3.36 mg/ml), the percentage chlorogenic acid concentration is 16.1% (wt). It is noted the two percentage concentrations obtained from the two methods are different. This is mainly because of the fact that concentrations of all compounds are measured against a single chlorogenic acid standard. Different spectroscopic properties of different compounds in different analytical conditions were not taken into account. Therefore, the equivalent concentrations here only provide a relative measure for the purpose of comparisons between results obtained from different extraction methods.

The chromatogram in Fig.2 showed that the extraction solution obtained from ethanol extraction contained a mixture of a number of polyphenol substances at relatively high concentrations. If the extraction solution is to be used as antibacterial agent, it may be of advantageous to have a mixture of polyphenol compounds. However, chlorogenic acid is one

of the components in the mixture, if pure chlorogenic acid is the final product required, the presence of other polyphenol compounds in high concentrations became undesirable. Although ethanol is the most commonly used solvent for chlorogenic acid extraction, it may not be the best solvent selection (16). The effectiveness of other extraction solvents can be studied to increase the relative concentration of the chlorogenic acid in the extraction solution.

Fig.3 shows the HPLC chromatogram of extraction solution from acetone water (1:2 v/v) extraction at 25 °C. As can be seen in Fig.3, the peak characteristics are quite different from those shown in Fig.2. In particular, the peak for chlorogenic acid is much larger than peaks for other compounds. Therefore, both relative and the absolute concentration of the chlorogenic acid were higher. Or, the purity of the chlorogenic acid in the extraction solution from the mixed solvent extraction was higher, indicating the mixed solvent has a higher selectivity for chlorogenic acid over other extracted compounds.

The chlorogenic acid concentration in Fig.3 was calculated to be 1.64 mg/ml, which represents 39.7% (peak area) of total concentrations from the HPLC method, or 46.1% (wt) of the total concentrations determined from the ferrous tartrate method. As the mixed solvent extraction was carried at 25 °C, the extraction time needed was long at 24 hr. As it was shown earlier, the extraction time can be much reduced by the use of higher extraction temperatures, or extraction aids such as ultrasound and microwave treatment.

Fig.4 is the HPLC chromatograms from extraction solutions from acetone water (1:2 v/v) extraction with ultrasound treatment. It can be seen that peak characteristics of the chromatograms in Fig. 4 are similar to those observed in Fig.3. Therefore, while the extraction time was much reduced with the use of ultrasound, the composition of the extract

solution, or the selectivity of the mixed solvent extraction for chlorogenic acid was not significantly changed by the ultrasound used. The chlorogenic acid concentration in Fig.4 was 2.12 mg/ml, which is 43.5% (peak area) of the total concentration from the HPLC method, or 48.0% (wt) of the total concentration from the ferrous tartrate method. It is noted here that the percentage concentrations are also similar to the case of mixed solvent extraction without the use of ultrasound.

Finally, the HPLC chromatogram of extraction solution from acetone water (1:2 v/v) extraction with microwave is shown in Fig.5. The peak characteristics as shown Fig.5 are also somewhat similar to those shown in Fig.3 and Fig.4. Or the use of microwave treatment did not greatly change the composition of the extraction solution. Similarly, the chlorogenic acid concentration in Fig.5 was 1.47 mg/ml, and the corresponding percentage concentrations were 34.5% (peak area) and 30.2% (wt), respectively. The percentage concentrations for the case of microwave treatment seemed to be lower than the case of ultrasound treatment.

## CONCLUSIONS

The extraction of chlorogenic acid and other polyphenol compounds from tobacco residues with the method of ethanol extraction, acetone water (1:2 v/v) mixed solvent extraction, with and without the use of ultrasound and microwave as the extraction aids were effectively achieved. The compositions of the various extraction solutions were also characterised by the use of HPLC method. The use of the extraction aids dramatically reduced the extraction time. The total amount of polyphenol compounds obtained from extraction experiments with the use of ultrasound and microwave were also significantly higher those obtained from simple solvent extractions.

Composition analysis results showed that the relative concentration, or the purity of the chlorogenic acid achieved from the acetone water (1:2 v/v) mixed solvent extraction was much higher than that from ethanol extraction, indicating better selectivity of the mixed solvent solution for chlorogenic acid. Overall, mixed solvent extraction with the use of ultrasound treatment achieved the best results in terms of both the relative and absolute concentrations chlorogenic acid of 48.0% (wt) and 2.12 mg/ml as determined from the HPLC method.

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TABLE 1. Total concentration of polyphenol compounds in extract solutions obtained with different extraction methods (tobacco residue: 10g; solvent: 90 ml)

Extraction method	Extraction temperature (°C)	Extraction time	Polyphenol concentration (mg/ml)
Ethanol extraction	78	2 hr	3.36
Acetone water(1:2v/v) extraction	25	2 day	3.55
Acetone water (1:2 v/v) ultrasound	25	15 min	4.42
Acetone water (1:2 v/v) microwave	25 <sup>a</sup>	90 sec	4.87

<sup>a</sup> microwave extraction temperature increased to 95°C.

## Figure Legends

Figure1. HPLC chromatogram of chlorogenic acid standard solution (2 mg/ml).

Figure 2. HPLC chromatogram of extraction solution from ethanol extraction at 78 °C (2 hr).

Figure 3. HPLC chromatogram of extraction solution from acetone water (1:2 v/v) extraction at 25 °C (2 days).

Figure 4. HPLC chromatogram of extraction solution from acetone water (1:2 v/v) extraction with ultrasound (15 min).

Figure 5. HPLC chromatogram of extraction solution from acetone water (1:2 v/v) extraction with microwave (90 sec).

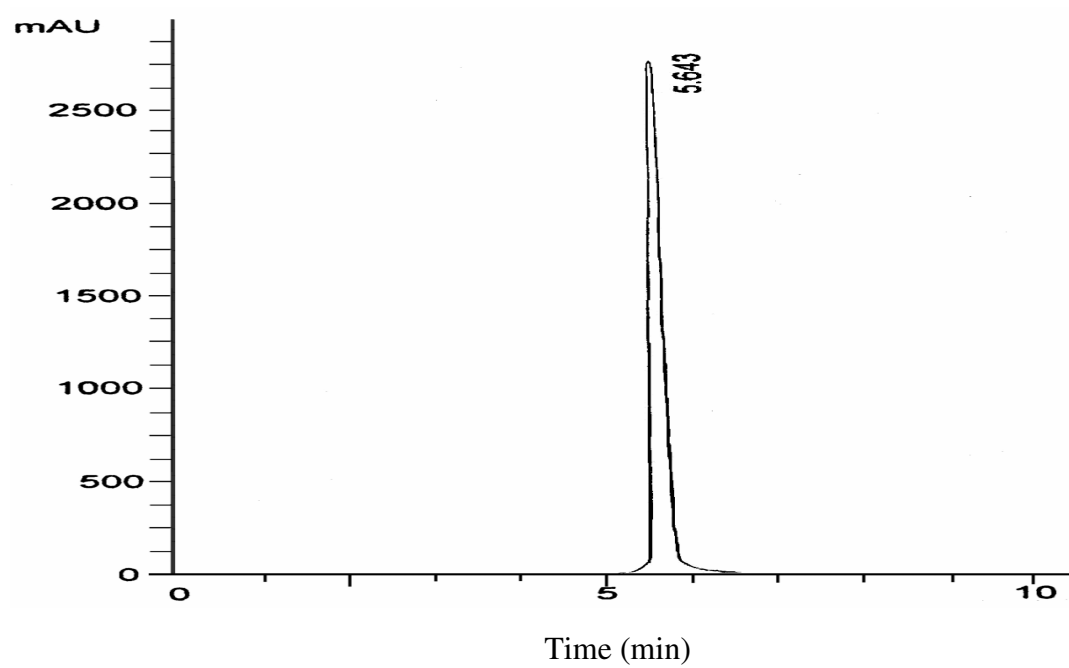


Figure 1

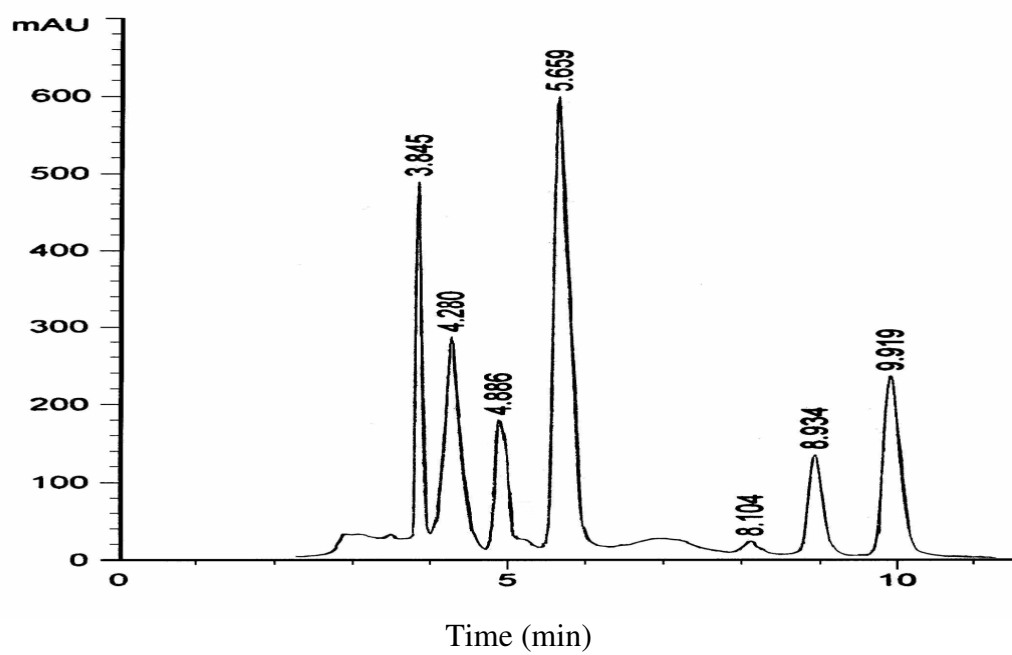


Figure 2

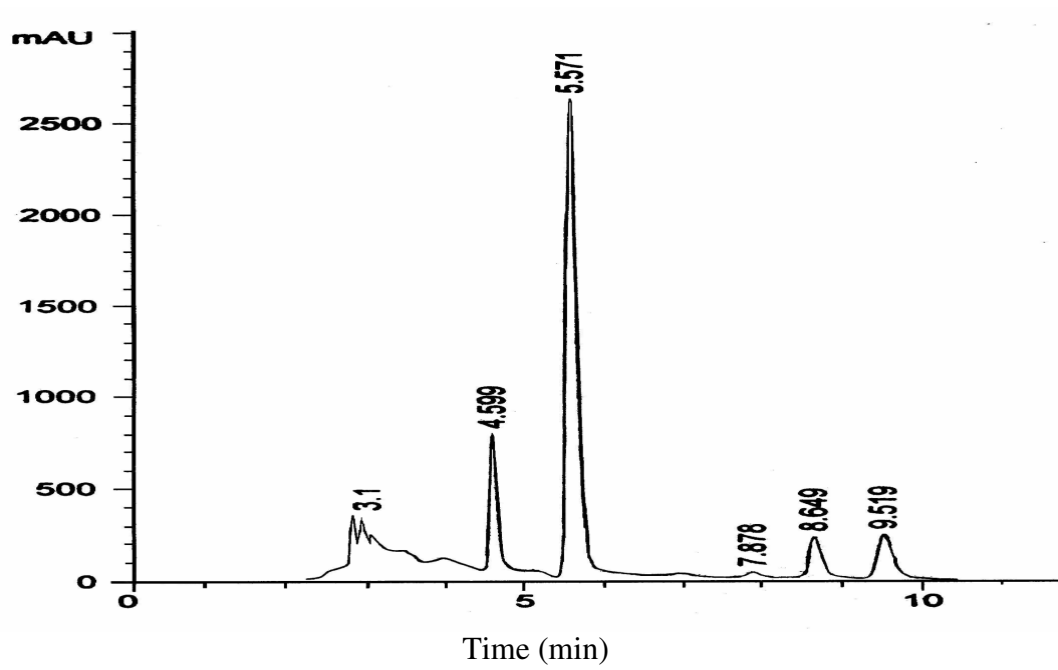


Figure 3

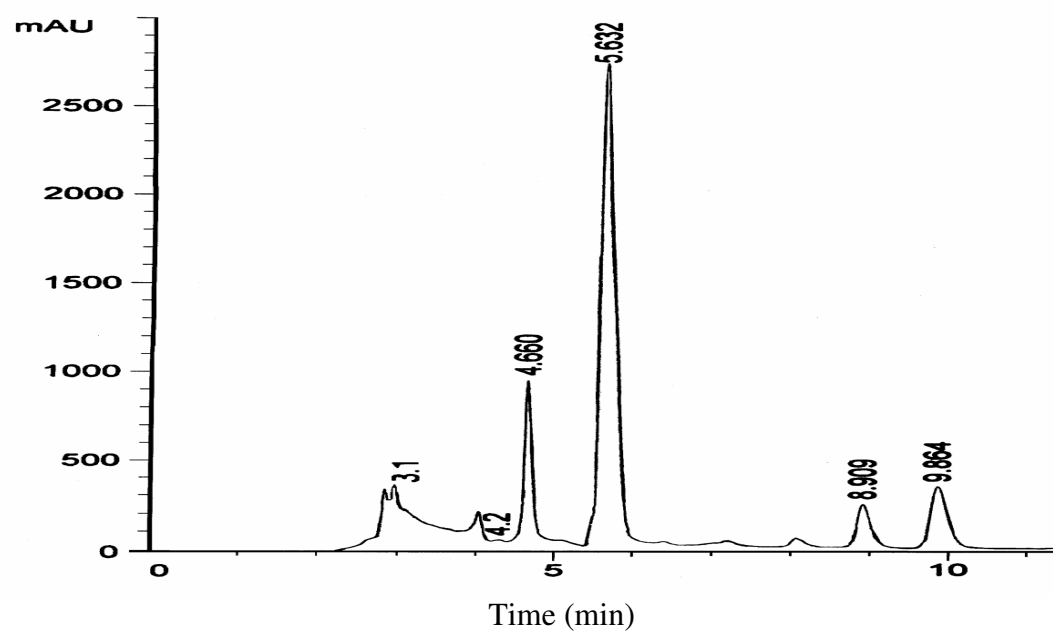


Figure 4

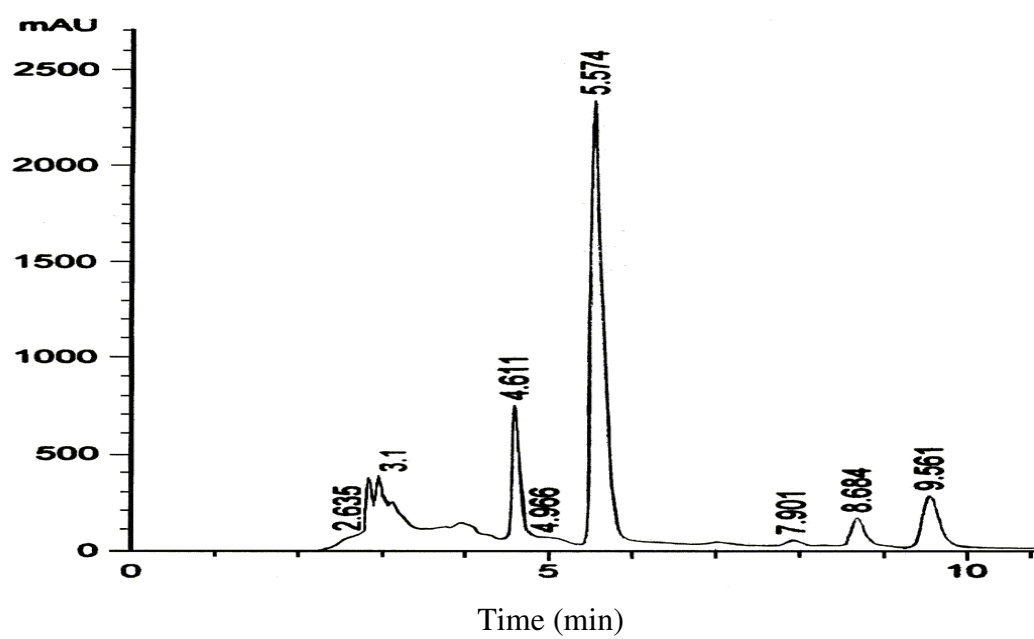


Figure 5