



The Phytochemistry and Chemotherapeutic Potential of *Tasmania lanceolata* (Tasmanian Pepper): A Review

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01 **Review Article** 0102
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05 **The phytochemistry and chemotherapeutic potential of** 05
06 ***Tasmannia lanceolata* (Tasmanian pepper): A review** 06
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10 1011 **Cock IE^{*,a,b}** 1112 ^aBiomolecular and Physical Sciences, Nathan Campus, Griffith University, 170 Kessels Rd, Nathan, Brisbane, Queensland 4111, Australia 1213 ^bEnvironmental Futures Centre, Nathan Campus, Griffith University, 170 Kessels Rd, Nathan, Brisbane, Queensland 4111, Australia 1314
15 **ABSTRACT:** Plants contain a myriad of natural compounds which exhibit important bioactive properties. These 15
16 compounds may provide alternatives to current medications and afford a significant avenue for new drug discovery. 16
17 Despite this, little information is available in the literature regarding many native Australian plants and their potential 17
18 for medicinal and industrial uses. *Tasmannia lanceolata* (Tasmanian pepper) has a long history of usage by Australian 18
19 Aborigines and European settlers as a food flavouring agent. Aborigines also used it for the treatment and cure 19
20 of skin disorders, venereal diseases, colic, stomach ache and as a quinine substitute. Apart from the reported 20
21 ethnopharmacological uses of Tasmanian pepper, surprisingly few studies have rigorously examined this species for its 21
22 medical properties. Recent studies have reported Tasmanian pepper to be an extremely good source of antioxidants. 22
23 Indeed, Tasmanian pepper has been reported to have free radical scavenging activities more than 4 times higher than 23
24 blueberries despite having ascorbic acid levels below the level of detection. Tasmanian pepper is particularly high in 24
25 terpenes and phenolic compounds but also has high levels of a variety of other antioxidants, including anthocyanins 25
26 and anthocyanins glycosides. Antioxidants have been associated with the prevention of cancer, cardiovascular disease 26
27 and neurological degenerative disorders. They are also linked with anti-diabetic bioactivities and have been associated 27
28 with the reduction of obesity. Antioxidants can directly scavenge free radicals, protecting cells against oxidative stress 28
29 related damage to proteins, lipids and nucleic acids. Therefore, *T. lanceolata* has potential in the treatment of a variety 29
30 of diseases and disorders and its potential bioactivities warrant further investigation. 3031 **KEYWORDS:** Winteraceae, *Tasmannia lanceolata*, Tasmanian pepper, Australian medicinal plants, antioxidants, 31
32 flavonoids, terpenoids 3233 **INTRODUCTION** 3334
35 Plants produce a wide variety of chemically diverse compounds which form the basis of their defense systems 35
36 against animal foraging, microbial infections and competition.^[1] These phytochemicals often have medicinally 36
37 important bioactivities and may be harnessed for new drug discovery or used directly as therapeutic agents. 37
38 Natural therapies not only form the basis of many traditional medicinal systems (particularly in developing 38
39 countries), but are also gaining wide spread acceptance and increased usage in Western medicinal systems. 39
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42 Furthermore, medicinal plants serve as a starting point 42
43 for natural products discovery and the development of 43
44 semi-synthetic agents with enhanced medical properties. 44
45 Indeed, approximately 25% of all prescription drugs cur- 45
46 rently in use were originally derived from plants or are 46
47 semi-synthetic analogues of plant derived compounds.^[2] 47
48 The statistics are even more impressive when we consider 48
49 the role of plants in the development of new anticancer 49
50 agents: approximately 75% of new anticancer drugs 50
51 marketed between 1981 and 2006 are derived from plant 51
52 compounds.^[1] Despite the impressive array of thera- 52
53 peutics derived from plants, only 10% of the estimated 53
54 250,000 species worldwide have been screened for any 54
55 bioactivities. Most of these studies have utilized tradi- 55
56 tional knowledge and ethnopharmacology to target spe- 56
57 cific plants. The study of plant pharmacognosy could 57
58 lead to the discovery of commercially and/or therapeuti- 58
59 cally useful phytochemicals possessing a diverse range of 59
60 activities. As Asian, Middle Eastern and European tradi- 60
61 tional medicine systems have been the most extensively 61
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documented compared to those of other world regions, the majority of studies have concentrated on plants from these regions. Recently there has been an increase in interest in the therapeutic potential of plants from other regions internationally and the medicinal potential of plants from Africa, South America and Australia are increasingly being reported.

As a result of its geographic isolation, Australia is home to a large variety of unique and distinct flora not found elsewhere in the world. Due to the harsh conditions seen in many parts of the continent, Australian plants have developed unique survival methods specific to the environmental conditions they inhabit and they may hold the key to the treatment of many diseases and medical conditions. Traditional Australian Aboriginal knowledge of plants as therapeutics is disappearing as the indigenous cultures merge into main stream society and the passing of oral traditions between each generation diminishes. Given the diverse nature of the flora present and the diminishing traditional knowledge, Australian plants remain relatively unstudied and it is surprising more research has not been done into their potential. A recent study into the antioxidant properties of several Australian plants has identified several species (including *Tasmannia lanceolata*) as being of particular interest due to their very high antioxidant activities and interesting phytochemistry.^[3,4]

The family Winteraceae

Winteraceae is a family of flowering plants consisting of approximately 90 species of trees and shrubs divided into 5 genera (*Drimys*, *Pseudowintera*, *Takhtajania*, *Tasmannia* and *Zygogynum*).^[5] The Winteraceae have developed as almost exclusively southern hemisphere plants, originating from precursor species on the Gondwana super continent. Their current distribution ranges from the cool climate regions of the southern Australia (particularly Tasmania) and New Zealand through to the temperate and tropical regions of Borneo, Madagascar, Molucca, New Caledonia, Papua New Guinea, the Philippines and Southern and Central America, with the majority concentrated in Australasia and Malesia.^[6]

The Winteraceae are characterized as woody evergreen plants with vessel-less xylem and plicate carpels.^[5,6] They generally have leaves without stipules. The leaves, which are almost always glabrous, have entire margins and are spirally arranged. Flowers are terminal, generally condensed and can be either bisexual or unisexual, depending on the individual species. The fruit forms as a fleshy berry with a hard seed. Many Winteraceae species are fragrant and are often used to produce essential oils. *Zygogynum*

is the largest Winteraceae genus with approximately 50 species.^[5-7] Until recently, *Belliolum*, *Bubbia* and *Exospermum* were classified as distinct genera, although most botanists now classify these as subgroupings of the *Zygogynum* genus. *Tasmannia* is the next largest genus with approximately 30 species.^[8] *Drimys* consists of 6 species, *Pseudowintera* has 2 species, and *Takhtajania* consists of a single species.^[8]

Members of family Winteraceae have been used for a broad range of dietary and medicinal purposes by a wide variety of ethnic and cultural groupings. The best documented of these is the South American species *Drimys winteri*. The stem and bark of this species has been used as a stimulant and as a tonic in traditional Brazilian medicinal systems.^[9] They are also used for the treatment of a wide variety of diseases and medicinal conditions including use as an analgesic, and to treat diarrhoea, inflammation, and ulcers.^[9,10] This species also has widespread usage in the treatment of scurvy due to its high antioxidant content.^[11] Of the other Winteraceae species, several have a history of ethnobotanical usage, usually for purposes related to their high antioxidant contents and as flavourants. Indeed, high levels of the compound polygodial (which gives the Winteraceae a characteristic peppery flavour) and high antioxidant contents are characteristic of several Winteraceae species.

Tasmannia lanceolata (Tasmanian pepper)

Tasmannia lanceolata (commonly known as Tasmanian pepper or mountain pepper; Figure 1a) is shrub which is endemic to the woodlands and cool temperate rainforests of Tasmania and the south-eastern region of the Australian mainland (Figure 1b). The species was originally described by the French botanist Jean Louis Poiret. Until 1969 it was classified in the genus *Drimys* and was named *Drimys lanceolata*. It is a medium to large shrub that varies between 2–5 m in height. Individual plants are unisexual, having either male or female flowers. The stems, branches and twigs are red in colour. The aromatic leaves are lanceolate to narrowly elliptical in shape (4–12 cm long, 0.7–2 cm wide) with a distinctly pale undersurface. Small creamy-white unisexual flowers appear during the summer months. These develop into small fleshy black 2 lobed berries (5–8 mm wide) during autumn.

As with many of the other Winteraceae species, *T. lanceolata* berries, leaves and bark have historical uses as a food and as a medicinal plant.^[12] When the berry is air dried it forms a small, hard peppercorn which is suitable for milling or crushing. The berry has a pleasant spicy flavor and sharp aroma. *T. lanceolata* was used as flavouring agent

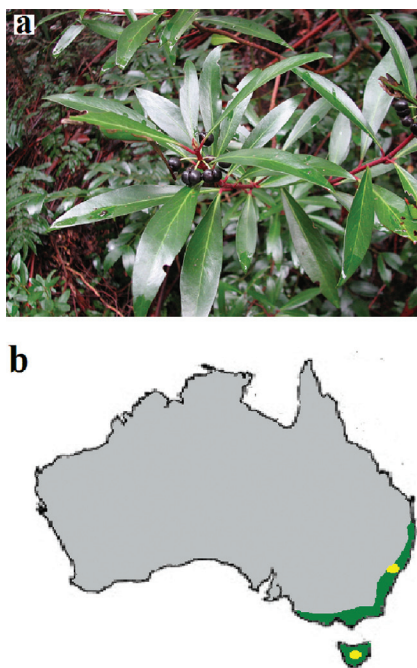


Figure 1. (a) *Tasmannia lanceolata* showing leaves and mature fruit. This photograph was obtained from Wikipedia commons (http://en.wikipedia.org/wiki/File:Tasmannia_lanceolata.jpg) and is reproduced here with the relevant permissions under the terms of the GNU Free Documentation Licence. (b) The distribution of *Tasmannia lanceolata* on the Australian continent (indicated by the green areas). The yellow areas indicate areas where the plant is particularly common.

by Australian Aborigines and more recently by European settlers. Historically, the leaves have been used as a herb and the berries have been used as a spice. Australian Aborigines also used *T. lanceolata* as a therapeutic agent to treat stomach disorders and as an emetic, as well as general usage as a tonic.^[13,14] Reports also exist of the use of *T. lanceolata* by Australian Aborigines for the treatment and cure of skin disorders, venereal diseases, colic, stomach ache and as a quinine substitute.^[13,15,16] Later, European colonists also recognized the therapeutic potential of *T. lanceolata* and the bark was used as a common substitute for other herbal remedies (including those derived from the related South American Winteraceae species, *Drimys wintera* (winter bark))^[17] to treat scurvy due to its high antioxidant activity.^[13,14]

ANTIOXIDANT CONTENT

Epidemiological studies have shown that a diet high in fruits/vegetables is associated with lower risk of developing chronic diseases.^[18] High antioxidant levels have previously been demonstrated to act as preventative effects

against the development of degenerative diseases such as cancer,^[19] cardiovascular diseases,^[20] neural degeneration,^[21] diabetes and obesity.^[22] The antioxidant activity of many plants has been associated with their phenolic contents. Many phenolic compounds have been shown to have strong antioxidant activities and may protect cells against oxidative damage by directly scavenging free radicals.^[23] Phenolic compounds may also interact directly with receptors or with enzymes involved in cellular signal transduction.^[24] Common classes of plant phenolic compounds include flavonoids, tannins and anthocyanins.

Recent studies have documented the exceptionally high antioxidant content of *T. lanceolata*.^[3,4] These studies have reported that *T. lanceolata* leaves have antioxidant contents more than 4 fold higher than those reported for blueberries (which themselves are considered to have high antioxidant contents). Interestingly, ascorbic acid (which itself makes a significant contribution to the antioxidant content of many fruits) was reported to be below the threshold of detection in this study and therefore would not contribute significantly to the high antioxidant content of *T. lanceolata*. Furthermore, the levels of *T. lanceolata* leaf phenolic antioxidants were reported in the same study to be over 3 fold higher than the levels in blueberries.^[4] *T. lanceolata* leaves have also been reported to have phenolic antioxidant contents up to 4 times higher than in basil leaves (*Ocimum basilicum*),^[25] higher levels than determined for peppermint leaves^[26] and similar levels to the phenolic antioxidant contents of maple, silver birch and spruce leaves.^[27] The antioxidant phenolic contents of *T. lanceolata* berries are also high, although these levels are significantly lower (less than 20%) than the leaf phenolic antioxidant levels. The contents are similar to those reported for those reported for *Piper nigrum* (black pepper) and *Lycium barbarum* (Chinese Barbary Wolfberry fruit),^[26] but approximately half the level of black sesame and peach kernel.^[27]

T. lanceolata leaves and berries also contains other compounds which contribute to their high antioxidant activities^[3,4] While many of these compounds are yet to be identified, *T. lanceolata* fruit has been shown to contain benzoic acids, flavanols, or flavanones.^[3] *T. lanceolata* is a good source of eugenol (Figure 2a), methyl eugenol (Figure 2b) and gallic acid (Figure 2c),^[28,29] all of which demonstrate strong antioxidant activity *in vitro*.^[30,31] *T. lanceolata* fruit extracts are also rich in lutein (Figure 2g—a carotenoid antioxidant compound associated with eye health) and with vitamin E (Figure 2g), vitamin A (Figure 2h) and folic acid (Figure 2i).^[3] The glycosides quercetin (Figure 2e) and rutin (Figure 2f) are some of the other

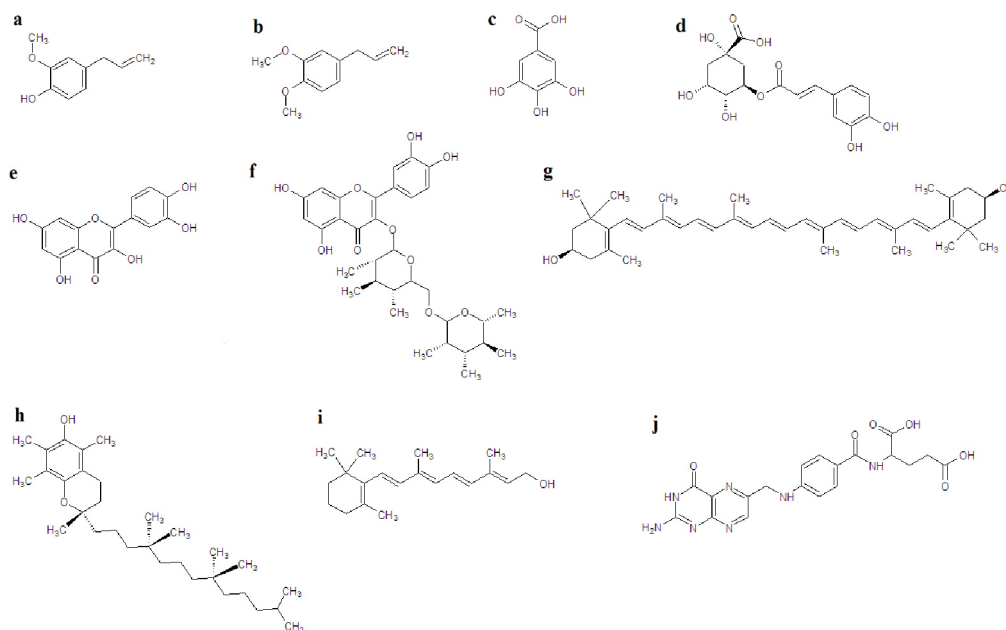


Figure 2. Chemical structures of selected antioxidant molecules identified in *T. lanceolata*: (a) eugenol, (b) methyl eugenol, (c) gallic acid, (d) chlorogenic acid, (e) quercetin, (f) rutin, (g) lutein, (h) α -tocopherol (vitamin E), (i) vitamin a, (j) folic acid.

antioxidants present in *T. lanceolata* fruit and leaves.^[3] *T. lanceolata* fruit is also a good source of the minerals magnesium, zinc, calcium, potassium, sodium, iron, phosphorous, manganese, copper, and molybdenum.^[3] It has previously been postulated that the exceptionally high antioxidant content of other plant species may be responsible for the therapeutic effects displayed by those plants.^[32,33] Therefore, it is likely that the high antioxidant contents reported for *T. lanceolata* extracts and essential oils would convey similar therapeutic properties.

The medicinal potential of plants with high antioxidant contents has been receiving much recent attention^[3,4] and reports have linked antioxidant levels and redox management with anticancer activity.^[32] A recent study has demonstrated that a fruit extract from a different plant rich in polyphenolic compounds (*T. ferdinandiana*) displayed anti-proliferative activity against a panel of cancer cell lines.^[34] Studies into the antioxidant/prooxidant effects of extracts from other plant species have demonstrated that the ability of a plant extract to exert antioxidant activity depends on multiple factors. Aloe vera antioxidant components for example may function as either antioxidants or prooxidants, with their action being dependent upon their concentration.^[33] The Aloe vera anthraquinone aloe emodin exerts antioxidant behaviour at lower concentrations, yet acts as a prooxidant at high concentrations. In contrast, a different Aloe vera anthraquinone (aloin) has an antioxidant effect at higher concentrations, yet a prooxidant effect at low concentrations. Thus, Aloe vera extracts and

components may act as either antioxidants or as oxidants, dependent on differing levels of the various constituents, and on their ratios. Thus, although *T. lanceolata* has very high antioxidant contents, it is possible that the individual components may act as either antioxidants or as oxidants and thus may also be effective in the treatment of cancer, as well as in its prevention at different concentrations.

Similar concentration dependent prooxidant effects have been reported for other antioxidant phytochemicals including many of the flavonoids^[35] which are present in high concentrations in *T. lanceolata* leaves and berries.^[36] Previous studies have also shown that the presence of transition metal ions such as copper or iron in the extract can enhance the conversion of the antioxidant to the prooxidant state.^[37,38] The prooxidant/antioxidant concentration dependent effects of plant extracts are due to a balance between the free radical scavenging activities and reducing power of their phytochemical components.^[32,33]

Reactive oxygen species (ROS) based tumour therapy would cause tumour regression should the tumour cells not be apoptotic/oxidant resistant cells. Therefore, if *T. lanceolata* antioxidant components are present in concentrations and ratios consistent with prooxidant activity, the extract would be expected to induce apoptosis and therefore would have anticancer activity. If the levels of components are consistent with a reducing environment, antioxidant activity would result and the extract would not have anticancer activity. Conversely, should the protocol

01 be repeated on a tumour with apoptotic resistant/oxidant
02 resistant cells, the converse would apply, where tumour
03 progression would be observed.

04 High antioxidant plants such as *T. lanceolata* also have
05 potential in the maintenance/control of diabetes.
06 Glycosylation of blood proteins including haemoglobin,
07 albumin and lipoproteins is characteristic of diabetes
08 mellitus.^[39] Under the hyperglycaemic conditions of dia-
09 betes mellitus, blood glucose interacts with specific amino
10 acids on the surface of proteins, forming glycosylated
11 protein products. These may undergo a series of further
12 chemical modifications, resulting in the production of
13 advanced glycation end products (AGE).^[40] The binding
14 of AGEs to their receptors results in altered cell signal-
15 ling which in turn results in free radical production.^[41]
16 Indeed, diabetes mellitus has been shown experimentally
17 to be associated with an increase in free radical formation
18 and an associated decrease in antioxidant potential ^[42,43]
19 Studies have directly linked oxidative stress with the
20 impaired maintenance of glucose homeostasis and the
21 enhanced lipid peroxidation seen in diabetes mellitus.^[42]
22 Furthermore, increased total antioxidant levels have been
23 measured in the blood and saliva of diabetic patients, fur-
24 ther supporting the proposed role of oxidative stress in
25 diabetes mellitus.^[44]
26

27 Oxidative stress induction has also been suggested to be
28 the common link between the diverse medical complica-
29 tions (including cardiovascular disease, renal and neural
30 degeneration, impaired vision and erectile dysfunction)
31 seen in diabetes mellitus.^[45,46] Therefore, treatment with
32 antioxidants would be expected to counteract many of
33 these complications. *T. lanceolata* leaves and berries have a
34 number of compounds (both phenolics and nonphenolic
35 compounds) that can act as antioxidants. Many phenolic
36 compounds could potentially behave as either antioxidant
37 or prooxidant dependant on their concentration, redox
38 state and ratio between compounds.^[33]
39

40 Eugenol (Figure 2a) has been shown to suppress the
41 growth of B16 melanoma and human HL-60 leukemia
42 cells.^[81] A recent study has also reported that eugenol
43 induces apoptosis in HCT-15 and HT-29 human colon
44 cancer cell lines.^[47] The same study also showed that
45 eugenol blocks cell cycle progression. Another study
46 reported that eugenol modulates cyclooxygenase 2
47 (COX-2) expression in HT-29 human colon cancer cells.^[48]
48 Furthermore, eugenol has additional therapeutic poten-
49 tial due to its other reported bioactivities.^[49] Its inges-
50 tion reduces the levels of blood glucose, triglycerides and
51 cholesterol, indicating its potential in the treatment and
52

01 maintenance of diabetes mellitus and as a hypolipidemic
02 agent. Eugenol relaxes arterial smooth muscle and has
03 potential as a vasodilator. It also has membrane stabiliz-
04 ing properties on synaptosomes, erythrocytes and mast
05 cells as well as providing it with therapeutic potential in
06 the treatment of inflammation and allergic disorders as
07 well as neurological conditions such as epilepsy. Eugenol
08 also has potential in the treatment of rheumatoid arthritis
09 due to its effect in lowering uric acid levels in rabbits.^[50] It
10 has also been reported to have antimicrobial activity.
11

12 PHYTOCHEMISTRY

13 *T. lanceolata* has been used as a flavouring agent by both
14 Aboriginal Australians as well as by later colonists and
15 settlers. It is well noted for its peppery taste and aroma.
16 Multiple studies have reported that the drimane sesqui-
17 terpene polygoidal (Figure 3a) is the major component
18 responsible for the flavour and aroma characteristics of
19 this species. Indeed, it has been reported that polygoidal
20 may account for nearly 40% of commercial *T. lanceolata*
21 essential oil components.^[51] Many studies have reported
22 the therapeutic properties of this compound, including
23 its antibacterial,^[52] antifungal,^[53–55] antihyperalgesia,^[56] anti-
24 inflammatory, antiallergic and vasorelaxation activities.^[57]
25
26

27 Studies examining the antibacterial activity of polygoi-
28 dal have provided conflicting reports. Early studies have
29 reported little or no antibacterial activity against limited
30 panels of bacteria, although many of these studies tested
31 polygoidal at relatively low concentrations (100 µg/ml).^[55]
32 In contrast, more recent studies have demonstrated
33 good bactericidal activity against both Gram-positive
34 and Gram-negative bacteria.^[52] Antifungal efficacy and
35 mechanistic studies have been more definitive, with
36 several publications highlighting polygoidal's potent
37 fungicidal activity.^[53–55] Polygoidal appears to exert its anti-
38 fungal activity by several mechanisms. It nonspecifically
39 disrupts/denatures fungal integral membrane proteins
40 by functioning as a nonionic surfactant.^[52] It also read-
41 ily reacts with amino acids (especially cysteine and aro-
42 matic amino acids), resulting in further denaturation. As
43 an additional antifungal mechanism, polygoidal may per-
44 meate cells by diffusing across the cell membrane. Once
45 inside the cell, polygoidal interacts with various intracel-
46 lular components and affects metabolic processes.
47

48 *T. lanceolata* also produces phenylpropenes including saf-
49 role (Figure 3b) and myristicin (Figure 3c). Similar phenyl-
50 propenes occur naturally in several other aromatic spices
51 including cinnamon, nutmeg, black pepper and basil.
52

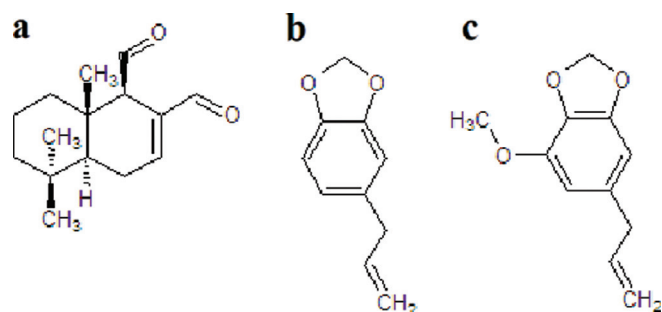


Figure 3. Chemical structures of (a) polygodial; (b) safrole; (c) myristicin.

The presence of safrole in *T. lanceolata* is concerning as it has been reported to be mildly genotoxic and carcinogenic in rats.^[58] Furthermore, safrole is also a weak hepatotoxin and has been shown to induce oxidative damage to liver cells.^[59] The carcinogenicity and toxicity of safrole has been shown to be due to the conversion by rat cytochrome P450 enzymes to electrophilic esters which form covalent adducts with DNA.^[60] In the past, safrole was widely used as an additive to beverages such as root beer and sassafras tea although its use is now banned by the US Food and Drug Administration (FDA) as a food additive and monitoring of its levels is recommended in products in which it occurs naturally. However, it must be noted that these early carcinogenesis/toxicity studies were performed in rodent experimental systems. Parallel safrole metabolism studies in humans demonstrated that the carcinogenic metabolites present in rat urine were absent in humans^[61] and thus the carcinogenic activity of safrole may be milder or even non-existent for humans. In contrast to safrole, the related compound myristicin (Figure 3c) has been reported to have tumorigenesis inhibitory activity via an induction of glutathione S-transferase activity.^[62]

Phenolics/flavonoids

Phenolic compounds, and in particular the flavonoids, have been identified as the major class of antioxidant compounds in *T. lanceolata*. As such, the phenolics have potential in the prevention and treatment of cancer and cardiovascular disease. Some flavonoids have been linked to the induction of cellular mechanisms that affect cancer cell progression and proliferation, as well as inhibiting tumour invasion.^[63] However, phenolics are also known to have further therapeutic properties in addition to their antioxidant activities (although some of these activities may be linked to the antioxidant activities). Flavonoids are considered to be particularly useful in maintaining good health and are often used as disease preventative agents. Preliminary reports suggest that flavonoids may modify

our responses to allergens, viruses and carcinogens.^[63] Indeed, studies have verified the antibacterial, antiviral, anti-inflammatory, anticancer and antidiarrhoeal activities of flavonoids.^[63]

Recent studies have reported very high levels of antioxidant flavonoids and flavonoid glycoside compounds in *T. lanceolata* extracts compared to the levels in other plants. These flavonoids include quercetin (Figure 2e), rutin (Figure 2f), (c) cyanidin-3-glucoside (Figure 4c) and cyaniding-3-rutinoside (Figure 4d). There is evidence that similar bioflavonoids prevent oxidation of LDL cholesterol via their free radical scavenging activity,^[64] inhibit endothelial activation^[65] and inhibit platelet aggregation.^[66] They also possess cyclooxygenase inhibitory activity and can prevent thrombosis.^[66] Evidence exists that the ingestion of high dietary levels of flavonoids is inversely proportional to the risk of coronary artery disease (CAD).^[67-69] It is therefore likely that the high flavonoid contents reported in *T. lanceolata* (particularly in the leaves) may have beneficial effects in CAD.

Recent studies have reported that many phenolic compounds also have potent anti-inflammatory activities.^[63] These anti-inflammatory effects are likely due to the inhibition of the enzymes cyclooxygenase and lipoxygenase, resulting in the inhibition of prostaglandin and leukotriene synthesis and the downstream release of cytokines.^[70,71] Quercetin (Figure 2e) in particular has been shown to

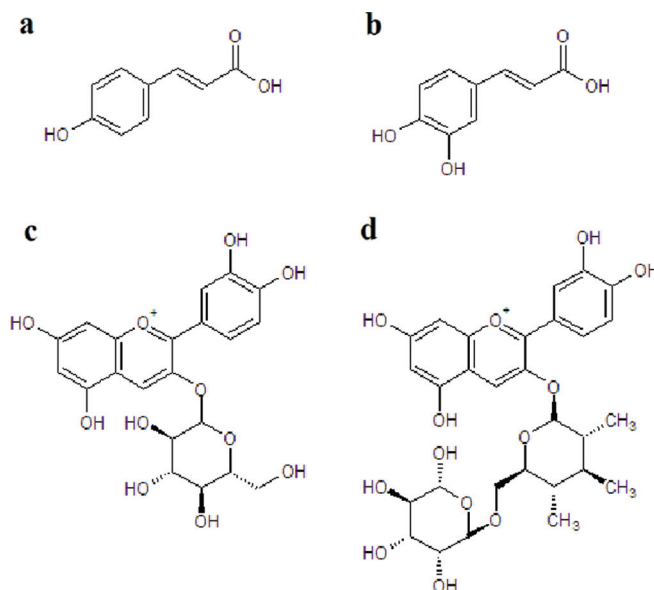


Figure 4. Chemical structures of known phenolic constituents of *T. lanceolata*: (a) coumaric acid, (b) caffeic acid, (c) cyanidin-3-glucoside, (d) cyanidin-3-rutinoside.

01 have potent inhibitory effects on both cyclooxygenase
02 and lipoxygenase enzyme activities via its antioxidant
03 activity, resulting in diminished eicosanoid biosynthesis.^[72]
04 These effects are exerted via a down regulation of cyclo-
05 oxygenase-2 (COX-2) and 5-lipoxygenase (5-LOX),
06 tumor necrosis factor-alpha (TNF- α) and interleukin-6
07 (IL-6).^[63] This down regulation results in the inhibition
08 of the inflammatory mediators such as nitric oxide (NO)
09 and prostaglandin E₂ (PGE₂) production. As mitogen-
10 activated protein kinases (MAPKs) which regulate inflam-
11 matory and immune responses may be activated by the
12 production of reactive oxygen species (ROS), it is likely
13 that the inhibition of ROS via quercetin is responsible for
14 its anti-inflammatory activity.

15
16 Whilst studies of the antibacterial activities of flavo-
17 noids vary widely (possibly due to intra and inter assay
18 variations), a number of flavonoids have been reported
19 to have antibacterial activity against multiple bacterial
20 species. One study examined the ability of quercetin and
21 rutin and their corresponding glycosides to inhibit the
22 growth of *Pseudomonas maltophilia* and *Enterobacter cloacae*.^[73]
23 This study showed that the quercetin glycosides showed
24 the strongest inhibitory activity of the flavonoids glyco-
25 sides tested. Many of the other glycosides also inhibited
26 bacterial growth, albeit with lower efficacy. Another study
27 tested the inhibitory activity of a panel of 38 flavonoids
28 against methicillin resistant *Staphylococcus aureus* (MRSA)
29 and reported moderate antibacterial activity for several
30 flavonoids including quercetin and luteolin. Rutin was
31 also shown to have a low MIC against multi-resistant
32 β -lactamase producing *Klebsiella pneumoniae*.^[74] Thus,
33 flavonoids have potential in the treatment of infective
34 diseases and much more study is required to examine the
35 structure/activity relationships of the compounds as well
36 as the mechanisms of their action.

37
38 Flavonoids also have antiviral bioactivities.^[68] Some of the
39 viral diseases that were reported to be inhibited by flavo-
40 noids were adenovirus, herpes viruses, HIV, parainfluenza
41 virus and respiratory syncytial virus.^[75,76] These studies
42 have shown that flavonoids have affects of on multiple
43 stages of viral replication and infectivity in vitro. For
44 example, quercetin exhibits both antiinfective and antirep-
45 licative bioactivities.^[75] Many of the current investigations
46 into the antiviral activities of flavonoids have reported on
47 their effects on the various stages of the HIV replicative
48 cycle. Most have focused on the ability of flavonoids to
49 inhibit HIV reverse transcriptase^[77] as well as antiintegrase
50 and antiprotease activities.^[68] Furthermore, epidemiologi-
51 cal studies have indicated that dietary flavonoids may have
52 a protective role against coronary disease.^[78]

Essential oil components

01 Volatile components account for the majority of the
02 *T. lanceolata* phytochemical profile, accounting for as high
03 as 6% of the dry weight of the plant material.^[51] For this
04 reason, until recently, research into *T. lanceolata* phytochem-
05 istry has largely concentrated on these components. A
06 recent analysis of commercial essential oil components^[51]
07 reported these to be predominantly sesquiterpenic, with
08 polygoidal (36.74%) (Figure 3a) being the major com-
09 ponent. Other sesquiterpenoids occur at lower levels in
10 *T. lanceolata* essential oils and are known to vary widely
11 between individual plants. An analysis of commercial
12 *T. lanceolata* essential oils^[51] reported that guaiol
13 (4.36%) (Figure 5f2), calamenene (3.42%) (Figure 5c2),
14 spathulenol (1.94%) (Figure 5f3), drimenol (1.91%)
15 (Figure 5c3), cadina-1,4-diene (1.58%) (Figure 5c1),
16 5-hydroxycalamenene (1.47%) (Figure 5d3), bicyclo-
17 germacrene (1.15%) (Figure 5e1), α -cubebene (0.88%)
18 (Figure 5c5), caryophyllene (0.87%) (Figure 5e3),
19 α -copaene (0.48%) (Figure 5c4), cadalene (0.44%)
20 (Figure 5b7), δ -cadinol (0.4%) (Figure 5d4), elemol
21 (0.39%) (Figure 5d2), T muurolol (0.39%) (Figure 5d5)
22 and germacrene D (Figure 5e2) are particularly abundant.
23 Other sesquiterpenoids present in *T. lanceolata* essential
24 oils include camphene (0.02%) (Figure 5d1), α -gurjunene
25 (0.04%) (Figure 5f1) and viridiflorol (Figure 5f4).^[51]
26

27
28 Several sesquiterpenes detected in *T. lanceolata* essen-
29 tial oils have been reported to have cytotoxic activities
30 against cancer cells. Polygoidal (the main component
31 of *T. lanceolata* essential oils) has demonstrated moder-
32 ate cytotoxicity towards V79 hamster lung fibroblasts,
33 Ehrlich ascites tumour cells (ECA) and mouse L1210
34 leukemia cell lines.^[79] That study also demonstrated
35 strong cytotoxic activity for drimenol and several of
36 its derivatives against a wide range of cancer cell lines.
37 β -caryophyllene induces apoptosis in PC-3 (prostrate
38 cancer) and MCF-7 (breast cancer) cell lines via ROS
39 mediated pathways.^[80] Similarly, β -caryophyllene and
40 camphene both demonstrate suppressive growth activ-
41 ity towards B16 melanoma and human HL-60 leukemia
42 cells.^[81] Cadalene and its derivatives (such as δ -cadinol)
43 inhibit lung tumourigenesis via the induction of apopto-
44 sis and by causing cell cycle arrest.^[82] T muurolol sesqui-
45 terpenoids have been shown to have mild cytotoxicity
46 towards several human tumour cell lines.^[83] Spathulenol
47 treatment blocks cell proliferation by inducing apoptosis
48 via caspase-3 independent pathways.^[84] *T. lanceolata* ses-
49 quiterpenoids have also been shown to block cell prolifer-
50 ation. Calamenene has been reported to exhibit potent
51 anti-proliferative activity against human A2780 ovarian
52 cancer cell lines.^[85]

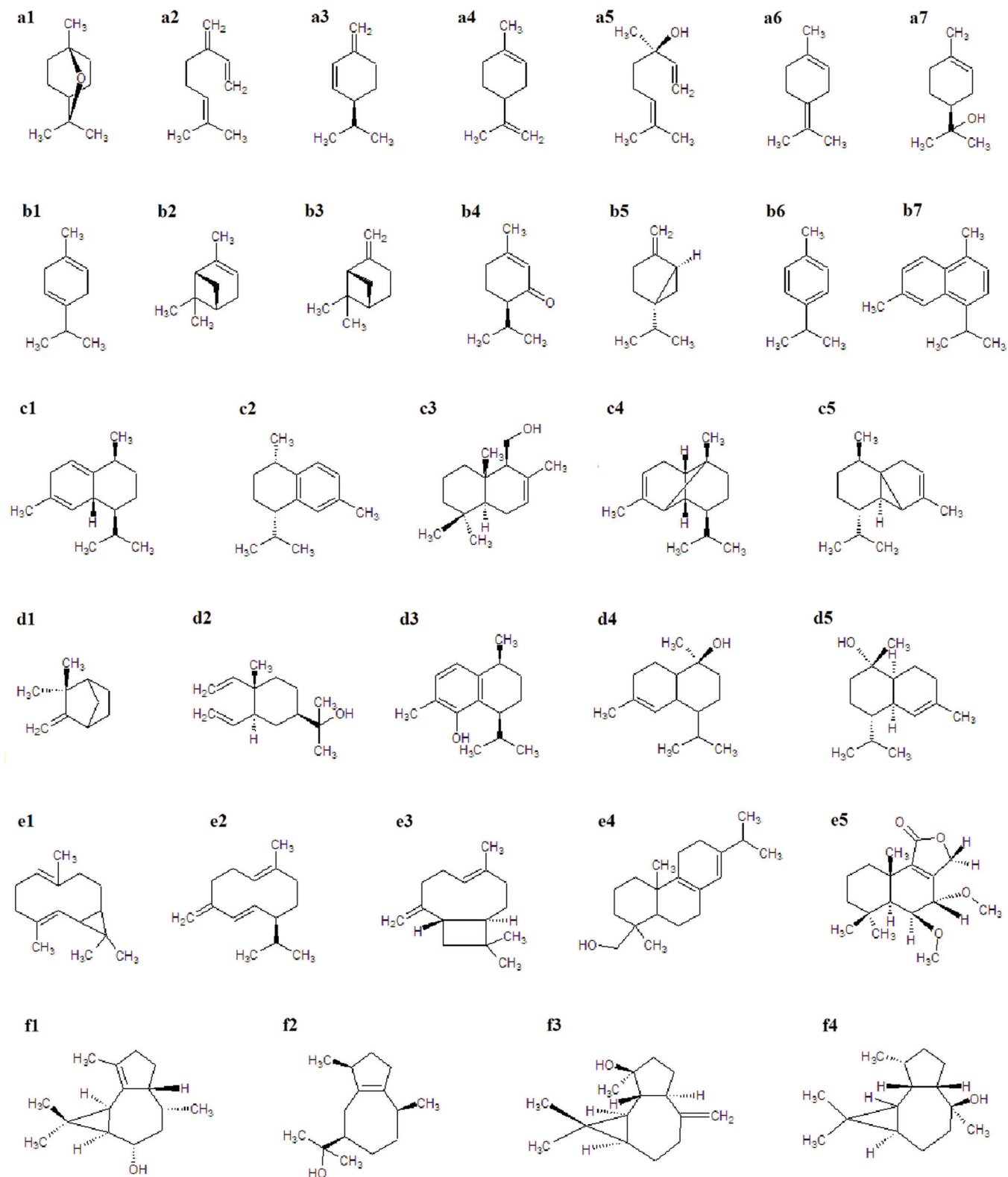


Figure 5. Chemical structures of terpenoid molecules identified in *T. lanceolata*: (a1) 1,8-cineole, (a2) myrcene, (a3) β -phellandrene, (a4) limonene, (a5) linalool, (a6) terpinolene, (a7) α -terpineol, (b1) γ -terpinene, (b2) α -pinene, (b3) β -pinene, (b4) piperitone, (b5) sabinene, (b6) cymene, (b7) cadalene, (c1) cadi-1,4-diene, (c2) calamenene, (c3) drimenol, (c4) α -copaene, (c5) α -cubebene, (d1) camphene, (d2) elemol, (d3) 5-hydroxycalamenene, (d4) δ -cadinol, (d5) T muurolol, (e1) bicyclgermacrene, (e2) germacrene D, (e3) caryophyllene, (e4) palustrol, (e5) drimenin, (f1) α -gurjunene, (f2) guaial, (f3) spathulenol, (f4) viridiflorol.

01 Many monoterpene compounds are also present in
02 significant levels in *T. lanceolata* with 1,8-cineole (0.77%)
03 (Figure 5a1), α -pinene (0.86%) (Figure 5b2), β -pinene
04 (0.38%) (Figure 5b3) and linalool (1.81%) (Figure 5a5)
05 predominating.^[51] Other characteristic monoterpenes
06 detected in the commercial *T. lanceolata* essential oils
07 analysed in that study included sabinene (Figure 5b5),
08 β -phellandrene (Figure 5a3), myrcene (Figure 5a2),
09 terpinolene (Figure 5a6), α -terpineol (Figure 5a7),
10 γ -terpinene (Figure 5b1), piperitone (Figure 5b4), limo-
11 nene (Figure 5a4) and cymene (Figure 5b6), although all
12 of these were generally present at levels below 0.1%.

13 Monoterpenes have been reported to exert a wide vari-
14 ety of biological effects including antibacterial, antifun-
15 gal, anti-inflammatory and antitumour activities. Several
16 monoterpenes detected in *T. lanceolata* essential oils have
17 been reported to have cytotoxic activities, directly killing
18 cancer cells. 1,8-cineol induces apoptosis in human leuka-
19 mia cell lines.^[86] Similarly, linalool induces apoptosis and
20 potentiates doxorubicin induced cytotoxicity in MCF-7
21 adenocarcinoma cell lines.^[87] Further studies have also
22 demonstrated that cotreatment of linalool with anthracy-
23 clines improves the therapeutic index in the management
24 of breast cancer cell lines.^[87] Pinene has been reported to
25 induce apoptosis in melanoma models.^[88] Several other
26 *T. lanceolata* essential oil monoterpene components also
27 display cytostatic activities against cancer cell lines. Limo-
28 nene is particularly promising as it blocks all phases of
29 cancer progression. Limonene has been shown to block
30 the induction of mammary cancer by 7, 12-dimethyl-
31 benzyl anthracene (DMBA).^[87] Furthermore, limonene also
32 blocks the progression of cancer post-initiation and is
33 effective in treating established breast cancers. In addition,
34 a comprehensive study examined the ability of a wide range
35 of terpenes to suppress the growth of B16 melanoma and
36 human HL-60 leukemia cells.^[81] Of the monoterpenes
37 previously reported to be present in *T. lanceolata* essen-
38 tial oils, 1,8-cineol, α -pinene, limonene, linalool, cymene,
39 α -terpineol and myrcene all were reported to have potent
40 tumour suppression activity in that study.

41 Several terpenoids have been reported to suppress
42 NF- κ B signaling (the major regulator of inflammatory
43 diseases and cancer).^[89] The monoterpenes limonene^[90,91]
44 and α -pinene^[92] have been reported to inhibit NF- κ B
45 signaling pathways. Limonene inhibition of mammary
46 and pancreatic tumours has been reported and has been
47 shown to be due to direct DNA binding.^[93] α -Pinene also
48 affects inflammatory diseases and cancer by inhibiting
49 p65 translocation into the nucleus in LPS-induced NF- κ B

01 signaling.^[92] Furthermore, many other sesquiterpenes
02 and sesquiterpene lactones also have well established
03 anticancer and anti-inflammatory activities.^[89] Whilst
04 much work is still needed to characterize the mechanisms
05 of action of these compounds, it appears that NF- κ B
06 inhibitory activities may be responsible.

07 The antimicrobial activity of *Drimys winteri* (a species
08 closely related to *T. lanceolata*) essential oils have been well
09 documented against a variety of bacterial species and it
10 has been established that terpenoids contribute to this
11 activity.^[94] *Drimys winteri* essential oils contain many of the
12 same monoterpene constituents as *T. lanceolata* essential
13 oils (including polygodial, α -pinene, β -pinene, sabinene,
14 myrcene, terpinene, limonene and β -phellandrene). That
15 study demonstrated good antibacterial activities for all
16 of these compounds. Further studies have also shown
17 that the monoterpene piperitone reduces the resistance
18 of several strains of Enterobacteriaceae to the antibacte-
19 rial agent nitrofurantoin.^[95] Other studies have reported
20 similar antibacterial activities for the sesquiterpenoids
21 α -cubebene, copaene and caryophyllene isolated from
22 *Pilgerodendron uviferum*.^[96]

23 Hydrocarbons

24 Unsaturated fatty acids and unsaturated hydrocarbons are
25 components in many plant oils including safflower oil, soy-
26 abean oil and cotton seed oils and have also been shown to
27 be abundant in *T. lanceolata* oils.^[51,97] Amongst these, lino-
28 lenic acid (Figure 6b) has received attention for its antioxi-
29 dant activity and therapeutic potential. Increased dietary
30 intakes of unsaturated fatty acids (including linolenic acid)
31 has been associated with a decreased incidence of cardio-
32 vascular disease.^[98] Linolenic acid has also been reported
33 to have anti-inflammatory activity due to its antioxidant
34 potential.^[99] The same study determined that linolenic acid
35 blocks nitric oxide synthase gene expression via NF- κ B
36 and mitogen activated protein kinase (MAPK) pathways,
37 resulting in inhibition of nitric oxide production. Thus it is
38 possible that linolenic acid may also have anticancer effects.
39 Similarly, squalene (Figure 6c) has therapeutic potential
40 and has been associated with the antioxidant activities of
41 other plant species.^[100] As squalene (Figure 6c) is known
42 to inhibit the ras gene,^[101] it is likely that it also affects can-
43 cer progression. Similarly, squalene inhibits inhibit HMG-
44 CoA reductase^[101] and thus it may lower endogenous sterol
45 synthesis and decrease cardiovascular disorders.

46 Medium length (C16-18) straight chain fatty acids (MCFA)
47 have been reported to have strong antimicrobial effects
48 against a wide variety of bacteria, fungi, viruses and

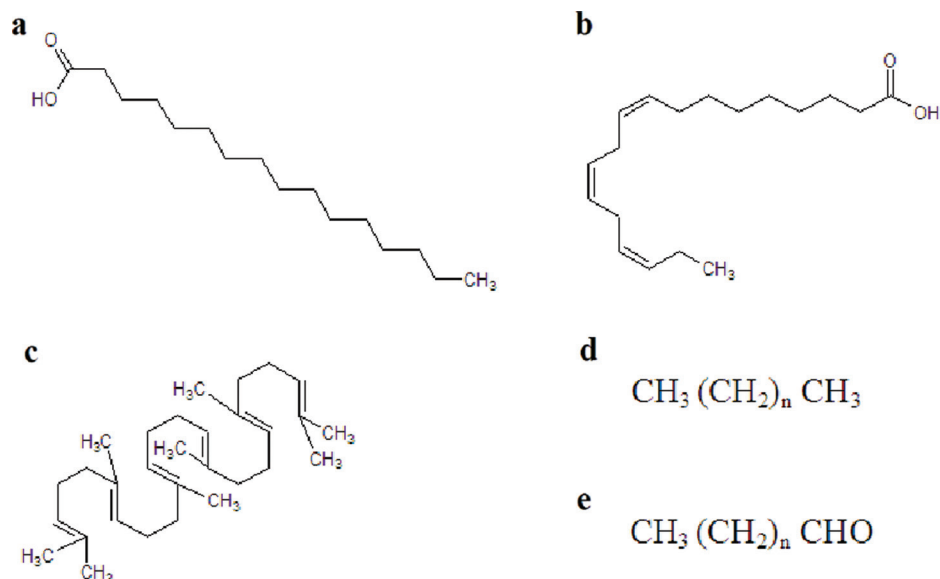


Figure 6. Chemical structures of selected hydrocarbon components identified in *T. lanceolata*: (a) palmitic acid, (b) linolenic acid, (c) squalene, (d) general alkane structure (common chain lengths detected in *T. lanceolata* extracts include C = 23, 25, 27), (e) general saturated primary fatty alcohols (common chain lengths detected in *T. lanceolata* extracts include C = 24, 26, 28).

protozoa. Multiple studies have reported the potential of MCFA in the control of such diverse pathogenic bacteria as *Bacillus anthracis*,^[102] *Neisseria gonorrhoeae*,^[103] *Heliobacter pylorus*,^[104] *Vibrio cholera*^[105] and various Streptococci species.^[106] MCFAs can also inactivate a wide range of infective viral agents including cytomegalovirus (CMV),^[107] Dengue virus,^[108] influenza,^[108] measles,^[108] polio virus,^[108] herpes viruses^[108] and HIV.^[110] Similarly, MCFA have been reported to have good fungicidal activity against the medicinally important fungi *Aspergillus niger*^[111] and *Candida albicans*^[112] and antiprotozoal activity against *Giardia duodenalis*.^[113] Of the MCFAs, the C18 straight chain unsaturated fatty acid linolenic acid (with is abundant in *T. lanceolata* extracts and essential oils^[51,97]) has been reported to have particularly potent antibacterial activity. Several reports have reported growth inhibition against *Bacillus cereus* and *Staphylococcus aureus* at concentrations as low as 10 µg/ml.^[114] More recently, linolenic acid has been reported to have antibacterial activity on its own against a broader range of bacteria, as well as increasing the antibacterial effects of monoglycerides.^[115] Of the other *T. lanceolata* fatty acids, the C16 straight chain saturated fatty acid palmitic acid has also been reported to have antibacterial activity against both Gram-negative and Gram-positive bacterial species.^[116] The same study also showed the ability of this MCFA to inhibit the replication of the influenza A virus.

The fatty alcohols and unbranched paraffins detected in *T. lanceolata* essential oils also have therapeutic potential. Both classes of compounds have surfactant properties.^[117]

Therefore they may nonspecifically disrupt/denature fungal integral membrane proteins and have potential as antibiotic agents. An increased intake of long chain fatty alcohols (C24-34) similar to those present in *T. lanceolata* extracts and essential oils^[51,97] has also been reported to lower LDL cholesterol levels by as much as 88%.^[118] Thus, it is possible that *T. lanceolata* ingestion may also have beneficial cardiovascular effects and more investigation is needed in this area.

CONCLUSION

Despite the history of traditional *T. lanceolata* usage, until recently, there has been little rigorous scientific research into the medicinal potential of this species. Recent studies,^[3,4] whilst initially focussed on the food properties of *T. lanceolata*, have also indicated the potential of this plant as a therapeutic agent. Indeed, several recent reports indicate a growing interest in examining medicinally important bioactivities induced by *T. lanceolata*. Recently, *T. lanceolata* has been reported to have good antioxidant,^[3,4] anticancer,^[119] antidiabetic^[120] and antimicrobial effects.^[121] In most cases the active phytochemicals have not been established although several of these studies have linked these activities to their antioxidant activities. Instead, often the partially purified compounds of a crude extract are itemised yet the active component(s) not identified. In other studies, the active compounds have not been characterised and instead only the classes of compounds in the crude mixture have been determined.

Given the impressive antioxidant activity of this species and the medicinal properties of many of its known phytochemicals, it is likely that bioactivity studies will detect further therapeutic properties for *T. lanceolata*. Much work is still required to fully understand the phytochemistry and pharmacognosy of *T. lanceolata*. Furthermore, few of these studies have provided substantial mechanistic detail to explain how the active principles achieve their medicinal effects.

Cancer is a major public health burden, both in developed and developing countries. Plant derived agents such as taxol, vinblastine, vincristine, and the camptothecin derivatives topotecan and irinotecan and etoposide (derived from epipodophyllotoxin) are in clinical use globally^[122] for the treatment of cancer. With regard to the phytochemical studies summarised in this review, it is surprising that the chemotherapeutic potential of *T. lanceolata* remains largely unexamined. Although *T. lanceolata* extracts and essential oils are not yet fully characterised due to difficulties in separating some components, high levels of antioxidant molecules have been reported. Apart from the antioxidant compounds discussed in this report, *T. lanceolata* also contains high levels of other phenolic and terpenoid compounds which have therapeutic potential that is not just limited to cancer treatment. Polar *T. lanceolata* extracts contains over 4-fold higher levels of antioxidants than in blueberries.^[3] Studies into the therapeutic potential of this species are still in their infancy and most of the studies regarding this plant are focussed on the total antioxidant capacity, with several recent studies beginning to examine the medicinally important bioactivities. The current review highlights the chemotherapeutic potential of the phytochemicals of *T. lanceolata*. In particular, this manuscript describes the potential of this plant in treatment for disorders related to cellular redox control (eg cellular proliferation, inflammation, cancer, diabetes, obesity, cardiovascular and neurodegenerative diseases).

REFERENCES

- Newman DJ, Cragg GM. Natural products as sources of new drugs over the last 25 years. *J Nat Prod.* 2007; 70:461–77.
- Newman DJ, Cragg GM, Snader KM. Natural products as sources of new drugs over the period 1981–2002. *J Nat Prod.* 2003; 66:1022–37.
- Konczak I, Zabarás D, Dunstan M, Aguas. Antioxidant capacity and hydrophilic phytochemicals in commercially grown Australian fruits. *Food Chem.* 2010; 123:1048–54.
- Netzel M, Netzel G, Tian Q, Schwartz S, Konczak I. Native Australian fruits – a novel source of antioxidants for food. *Innov Food Sci Emerg Technol.* 2007; 8:339–46.
- Marquinez X, Lohmann LG, Faria Salatino ML, Salatino A, González F. Generic relationships and dating of lineages in Winteraceae based on nuclear (ITS) and plastid (*rpS16* and *psbA-trnH*) sequence data. *Mol Phylogenet Evol.* 2009; 53:435–49.
- Vink W. Taxonomy of Winteraceae. *Taxon.* 1988; 37:691–8.
- Mabberley DJ (2008) *Mabberley's Plant-Book. A portable dictionary of plants, their classification and uses.* 3rd edn. Cambridge: Cambridge University Press.
- Field TS, Zwieniecki MA, Holbrook NM. Winteraceae evolution: An ecophysiological perspective. *Ann Missouri Bot Gard.* 2000; 87(3):323–34.
- Correa MP. *Diccionario das plantas uteis do Brasil e das exóticas cultivadas.* Rio de Janeiro: Imprensa Nacional. 1984; 2
- Cunha FM, Frode TS, Mendes GL, Malheiros A, Cechinel Filho V, Yunes RA, Calixto JB. Additional evidence for the anti-inflammatory and anti-allergic properties of the sesquiterpene polygodial. *Life Sci.* 2001; 70:159–69.
- Bown D. *New Encyclopaedia of Herbs and their Uses: The Herb Society of America.* DK Publishing. 2001: London, UK.
- Gorman JT, Griffith AD, Whitehead PJ. An Analysis of the use of Plant Products for Commerce in Remote Aboriginal Communities of Northern Australia. *Econ Bot.* 2006; 60:362–73.
- Cock IE. Medicinal and aromatic plants – Australia, in *Ethnopharmacology section, Biological, Physiological and Health Sciences, Encyclopedia of Life Support Systems (EOLSS)*, 2011; Developed under the Auspices of the UNESCO, EOLSS Publishers, Oxford, UK, (<http://www.eolss.net>).
- Lassak EV, McCarthy T. *Australian Medicinal Plants. A complete guide to identification and usage.* Reed New Holland Publishers 2011, Sydney Australia.
- Retamar JA. *Essential Oils from Aromatic Species.* In 'On essential oils', 1986 edited by James Verghese. Kolenchery, India: Synthite; pp. 123–280.
- Salmon JT. *The native trees of New Zealand.* 1980, Reed Auckland.
- LeStrange R. *A history of herbal plants.* 1977, Angus and Robertson.
- Potter JD. Cancer prevention: epidemiology and experiment. *Cancer Lett.* 1997; 114:7–9.
- Hertog MG, Bueno-de-Mesquita HB, Fehily AM, Sweetnam PM, Elwood PC, Kromhout D. Fruit and vegetable consumption and cancer mortality in the caerphilly study. *Cancer Epidemiol Biomarkers Prev.* 1996; 5:673–7.
- Vita JA. Polyphenols and cardiovascular disease: Effects on endothelial and platelet function. *Am J Clin Nutr.* 2005;81(1):292S–7S.
- Youdim KA, Spencer JPE, Schroeter H, Rice-Evans CA. Dietary flavonoids as potential neuroprotectants. *Biol Chem.* 2002; 383:503–19.
- Tsuda T, Horio F, Uchida K, Aoki H, Osawa T. Dietary cyanidin 3-O-b-D-glucoside-rich purple corn colour prevents obesity and ameliorates hyperglycemia in mice. *J Nutr.* 2003; 133:2125–30.
- Rice-Evans C, Miller N, Paganga. Antioxidant properties of phenolic compound. *Trends Plant Sci.* 1997; 2(4):152–9.
- Moskaug JO, Carlsen H, Myhrstad MC, Blomhoff R. Polyphenols and glutathione synthesis regulation. *Am J Clin Nutr.* 2005; 81:277–83S.
- Javanmardi J, Stushnoff C, Locke E, Vivanco JM. Antioxidant activity and total phenolics content of Iranian *Ocimum accessions*. *Food Chem.* 2003; 83:547–50.
- Liu H, Qiu N, Ding H, Yao R. Polyphenols content and antioxidant capacity of 68 Chinese herbals suitable for medicinal and food uses. *Food Res Int.* 2008; 41:363–70.
- Kahkonen MP, Hopia AI, Vuorela HJ, Rahua JP, Pihlaja K, Kujala TS, et al. Antioxidant activity of plant extracts containing phenolic compounds. *J Agr Food Chem.* 1999; 47:3954–62.
- Cunningham AB, Garnett S, Gorman J, Courtenay K, Boehme D. Eco-Enterprises and *Terminalia ferdinandiana*: "Best Laid Plans" and Australian Policy Lessons. *Econ Bot.* 2009; 63:16–28.
- Cherikoff V, Kowalski G. Superfoods for Superhealth. Discover the Wonders of Australian Native Fruits. 2008. [Last cited on 2010 Oct 07]. Available from: http://www.kakadujuice.com/clientinc/upload/Promotional_Tools/SuperFoods%20Excerpt.pdf.
- Losso JN, Bansode RR, Trappey A, Bawadi HA, Truax R. In vitro antiproliferative activities of ellagic acid. *J Nutr Biochem.* 2004; 15:672–78.
- Ohno Y, Fukuda K, Takemura G, Toyota M, Watanabe M, Yasuda N, et al. Induction of apoptosis by gallic acid in lung cancer cells. *Anticancer Drugs.* 1999; 10:845–51.
- Mohanty S, Cock IE. The chemotherapeutic potential of *Terminalia ferdinandiana*: Phytochemistry and bioactivity. *Pharmacog Rev.* 2012; 6(11):29–36.
- Cock IE. Problems of reproducibility and efficacy of bioassays using crude extracts, with reference to Aloe vera. *Pharmacog Commns.* 2011; 1(1):52–62.

- 01 34. Tan AC, Konczak I, Ramzan I, Sze DMY. Native Australian fruit polyphenols inhibit cell viability and induce apoptosis in human cancer cell lines. *Nutr Cancer*. 2011; 63(3):444–55.
- 02 35. Rahman A, Shahabuddin M, Hadi SM, Parish J. Complexes involving quercetin, DNA and Cu(II). *Carcinogenesis*. 1990; 11:2001–3.
- 03 36. Ahsan H, Hadi SM. Strand Scission in DNA induced by curcumin in the presence of Cu(II), *Cancer Lett*. 1998; 124:23–30.
- 04 37. Lastra CA, Villegas I. Resveratrol as an antioxidant and pro-oxidant agent: Mechanisms and clinical implications, *Biochem Soc T*. 2007; 35:1156–60.
- 05 38. Wu CC, Lu YH, Wei BL, Yang SC, Won SJ, Lin CN. Phloroglucinols with prooxidant activity from *Garcinia subelliptica*. *J Nat Prod*. 2008; 71:246–50.
- 06 39. Klein R. Hyperglycemia and microvascular and macrovascular disease in diabetes. *Diabetes Care*. 1995; 18:258–68.
- 07 40. Singh R, Barden A, Mori, T, Beilin L. Advanced glycation end-products: a review. *Diabetologia*. 2001; 44:129–46.
- 08 41. Penckofer S, Schwertz D, Florczak K. Oxidative stress and cardiovascular disease in type 2 diabetes: the role of antioxidants and prooxidants. *J Cardiovasc Nurs*. 2002; 16(2):68–85.
- 09 42. Davi G, Ciabattini G, Consoli A, Mezzetti A, Falco A, Santarone S, Pennese E, Vitacolonna E, Bucciarelli T, Constantini F, Capani F, Patrono C. 1999. In vivo formation of 8-iso-prostaglandin F2a and platelet activation in diabetes mellitus: effects of improved metabolic control and vitamin E supplementation. *Circulation*. 1999; 99:224–9.
- 10 43. Vessby J, Basu S, Mohsen R, Berne C, Vessby B. Oxidative stress and antioxidant status in type 1 diabetes mellitus. *J Intern Med*. 2002; 251:69–76.
- 11 44. Astaneie F, Afshari M, Mojtahedi A, Mostafalou S, Zamani MJ, Larjani B, Abdollah M. Total antioxidant capacity and levels of epidermal growth factor and nitric oxide in blood and saliva of insulin-dependent diabetic patients. *Arch Med Res*. 2005; 36(4):376–81.
- 12 45. Rahimi R, Nikfar S, Larjani B, Abdollahi M. A review on the role of antioxidants in the management of diabetes and its complications. *Biomed Pharmacother*. 2005; 59:365–73.
- 13 46. Shih CC, Wu YW, Lin WC. Antihyperglycemic and antioxidant properties of *Anoectochilus Formosanus* in diabetic rats. *Clin Exp Pharmacol P*. 2002; 29:684–8.
- 14 47. Jaganathan SK, Mazumdar A, Mondhe D, Mandal M. Apoptotic effect of eugenol in human colon cancer cell lines. *Cell Biol Int*. 2011; 35:607–15.
- 15 48. Kim SS, Oh OJ, Min HY, Park EJ, Kim YG, Park HJ, Han YN, Lee SK. Eugenol suppresses cyclooxygenase-2 expression in lipopolysaccharide-stimulated mouse macrophage RAW264.7 cells. *Life Sci*. 2003; 73:337–48.
- 16 49. Kamatou GP, Vermaark I, Viljoen AM. Eugenol – From the remote Maluku Islands to the international marketplace: A review of a remarkable and versatile molecule. *Molecules* 2012; 17:6953–81.
- 17 50. Sarkar A, Pandey DN, Pant MC. A report on the effect of *Ocimum sanctum* (Tulsi) leaves and seeds on blood and urinary uric acid, urea and urine volume in normal albino rabbits. *Indian J Physiol Pharmacol*. 1990; 34:61–2.
- 18 51. Menary RC, Dragar VA, Thomas S, Read CD. Mountain pepper extract. *Tasmannia lanceolata*. Quality stabilisation and registration. Rural Industries research and development Corporation (RIRDC). 2003; Publication number 02/148.
- 19 52. Kubo I, Fujita K, Lee SH, Ha TJ. Antibacterial activity of polygodial. *Phytother Res*. 2005; 19:1013–17.
- 20 53. De Almeida Alves TM, Ribeiro FL, Kloos H, Zani CL. Polygodial, the fungitoxic component from the Brazilian medicinal plant *Polygonum punctatum*. *Mem Inst Oswaldo Cruz*. 2001; 96(6):831–3.
- 21 54. Lee SH, Lee JR, Lunde CS, Kubo I. 1999. In vitro antifungal susceptibilities of *Candida albicans* and other fungal pathogens to polygodial, a drimane sesquiterpene dialdehyde. *Planta Med*. 1999; 65:204–8.
- 22 55. Taniguchi M, Adachi T, Oi S et al. Structure-activity relationship of the Warburgia sesquiterpene dialdehydes. *Agric Biol Chem*. 1984; 48:73–8.
- 23 56. Mendes GL, Santos ARS, Campos MM et al. Antihyperalgesic properties of the extract and of the main sesquiterpene polygodial isolated from the barks of *Drymis winteri* (Winteraceae). *Life Sci*. 1998; 63:369–81.
- 24 57. Da Cunha FM, Frode TS, Mendes GL, Malheiros A, Cechinel-Filho V, Yunes RA, Calixto JB. Additional evidence for the anti-inflammatory and anti-allergic properties of the sesquiterpene polygodial *Life Sci*. 2001; 70:159–69.
- 25 58. Miller EC, Sxanson AB, Phillips DH, Fletcher TL, Liem A. Structure-activity studies of the carcinogenicities in the mouse and rat of some naturally occurring and synthetic alkybenzene derivatives related to saffrole and estragole. *Cancer Res*. 1981; 43:1124–34.
- 26 59. Liu TY, Chen CC, Chen CL, Chi CW. Saffrole-induced oxidative damage in the liver of Sprague-Dawley rats. *Food Chem Toxicol*. 1999; 37:697–702.
- 27 60. Miller JA, Miller EC. The metabolic activation and nucleic acid adducts of naturally-occurring carcinogens. Recent results with ethyl carbamate and the spice flavors saffrole and estragole. *Brit J Cancer*. 1983; 48:1–15.
- 28 61. Strolin Benedetti M, Mainoe A, Louis Broillet A. Absorption, metabolism and excretion of saffrole in the rat and man. *Toxicol*. 1977; 7:69–83.
- 29 62. Zheng G, Kenney PM, Zhang J, Lam LKT. Inhibition of benzo[a]pyrene-induced tumorigenesis by myristicin, a volatile aroma constituent of parsley leaf oil. *Carcinogenesis*. 1992; 13(10):1921–3.
- 30 63. Nijveldt RJ, van Nood, van Hoorn DEC, Boelens PG, van Norren K, van Leeuwen PAM. Flavonoids: A review of probable mechanisms of action and potential applications. *Am J Clin Nutr*. 2001; 74:418–25.
- 31 64. Fuhrman B, Aviram M. Antiatherogenicity of nutritional compounds. *The Invest Drug J*. 2001; 4:82–92.
- 32 65. Carluccio MA, Sicuella L, Ancora MA, Massaro M, Scoditti E, Storelli C, Visioli F, Distante A De Caterina R. Olive oil and red wine antioxidant polyphenols inhibit endothelial activation: antiatherogenic properties of Mediterranean diet phytochemicals. *Arterioscl Thromb Vas*. 2003; 23:622–9.
- 33 66. Ruff JC. Wine and polyphenols related to platelet aggregation and atherothrombosis. *Drug Exp Clin Res*. 2003; 25:125–31.
- 34 67. Martikainen JA, Ottelin AM, Kiviniemi V, Gylling H. Plant stanol esters are potentially cost-effective in the prevention of coronary heart disease in men: Bayesian modelling approach. *Eur J Cardiovasc Prevent Rehab*. 2007; 14:265–72.
- 35 68. Middleton EJ. Effect of plant flavonoids on immune and inflammatory cell function. *Adv Exp Med Biol*. 1998; 439:175–82.
- 36 69. Peluso MR. Flavonoids attenuate cardiovascular disease, inhibit phosphodiesterase and modulate lipid homeostasis in adipose tissue and liver. *Exp Biol*. 2006; 231:1287–99.
- 37 70. Ferrandiz ML, Alcaraz MJ. Anti-inflammatory activity and inhibition of arachidonic acid metabolism by flavonoids. *Agents Actions*. 1991; 32:283–8.
- 38 71. Laughton MJ, Evans PJ, Moroney MA, Hoult JR, Halliwell B. Inhibition of mammalian 5 lipoxygenase and cyclo-oxygenase by flavonoids and phenolic dietary additives. Relationship to antioxidant activity and to iron ion-reducing ability. *Biochem Pharmacol*. 1991; 42:1673–81.
- 39 72. Kim HP, Mani I, Iversen L, Ziboh VA. Effects of naturally-occurring flavonoids and bioflavonoids on epidermal cyclooxygenase and lipoxygenase from guinea-pigs. *Prostag Leukotr Ess* 1998; 58: 17–24.
- 40 73. Waage SK, Hedin PA. Quercetin 3-O-galactosyl-(1Æ6)-glucoside, a compound from narrowleaf vetch with antibacterial activity. *Phytochem*. 1985; 24:243–5.
- 41 74. Özcelik B, Deliorman Orhan D, Özgen S, Ergun F. Antimicrobial activity of flavonoids against extended-spectrum β-lactamase (ESβL)-producing *Klebsiella pneumoniae*. *Trop J Pharm Res*. 2008; 7(4):1151–7.
- 42 75. Kaul TN, Middleton E Jr, Ogra PL. Antiviral effect of flavonoids on human viruses. *J Med Virol*. 1985; 15:71–9.
- 43 76. Wang HK, Xia Y, Yang ZY, Natschke SL, Lee KH. Recent advances in the discovery and development of flavonoids and their analogues as antitumor and anti-HIV agents. *Adv Exp Med Biol*. 1998; 439:191–225.
- 44 77. Ng TB, Huang B, Fong WP, Yeung HW. Anti-human immunodeficiency virus (anti-HIV) natural products with special emphasis on HIV reverse transcriptase inhibitors. *Life Sci*. 1997; 61:933–49.
- 45 78. de Groot H, Rauhen U. Tissue injury by reactive oxygen species and the protective effects of flavonoids. *Fundam Clin Pharmacol*. 1998; 12:249–55.
- 46 79. Jansen BJM, de Groot AE. Occurrence, biological activity and synthesis of drimane sesquiterpenoids. *Nat Prod Rep*. 2004; 21: 449–77.
- 47 80. Park KR, Nam D, Yun HM, Lee SG, Jang HJ, Sethi G, Cho SK, Ahn KS. β-Caryophyllene oxide inhibits growth and induces apoptosis through the suppression of PI3K/AKT/mTOR/S6K1 pathways and ROS-mediated MAPKs activation. *Cancer Letters*. 2011; 312:178–88.
- 48 81. Tatman D, Mo H. Volatile isoprenoid constituents of fruits, vegetables and herbs cumulatively suppress the proliferation of murine B16 melanoma and human HL-60 leukemia cells. *Cancer Lett*. 2002; 175:129–39.
- 49 82. Jin H, Kim HW, Xu CX, Kwon JT, Hwang SK, Lee ES, Chang SH, Park SJ, Noh MS, Woo MA, Yu KM, Lee HJ, Choi JW, Choi DH, Cho MH. Effects of 7-hydroxy-3-methoxycadalenone on cell cycle, apoptosis and protein translation in A549 lung cancer cells. *Biofactors*. 2007; 29:67–75.
- 50 83. Ding L, Pfoh R, Rühl S, Qin S, Laatsch H. T-muurolol sesquiterpenes from the marine *Streptomyces* sp. M491 and revision of the configuration of previously reported amorphanes. *J Nat Prod*. 2009; 72:99–101.

84. Ziaei A, Ramezani M, Wright L, Paetz C, Schneider B, Mairghofran Z. Identification of spathulenol in *Salvia mirzayanii* and the immunomodulatory effects. *Phytotherapy Research* 2010; 25:557–62.
85. Dai Y, Harinantenaina L, Brodie PJ, Callmander M, Randrianasolo S, Rakotobe E, Rasamison VE, Kingston DGI. Isolation, synthesis and bioactivity of Calamenene sesquiterpenoids from *Sterculia capuronii* from Madagascar dry forest. *Planta Med.* 2012; 78:CL16.
86. Moteki H, Hibasami H, Yamada Y, Katsuzaki H, Imai K, Komiya T. Specific induction of apoptosis by 1,8-cineol in two human leukemia cell lines, but not in human stomach cancer cell line. *Oncol Rep.* 2002; 9(4):757–60.
87. Gould MN. Cancer chemoprevention and therapy by monoterpenes. *Environ Health Persp.* 1997; 105(4):977–9.
88. Matsuo AL, Figueiredo CR, Arruda DC, Pereira FV, Borin Scutti JA, Massaoka MH, Travassos LR, Sartorelli P, Lago JHG. A-Pinene isolated from *Schinus terebinthifolius* Raddi (Anacardiaceae) induces apoptosis and confers antimetastatic protection in a melanoma model. *BBRC.* 2011; 411:449–54.
89. Salminen A, Lehtonen M, Suuronen T, Kaarniranta K, Huuskonen J. Terpenoids: Natural inhibitors of NF- κ B signalling with anti-inflammatory and anticancer potential. *Cell Molec Life Sci.* 2008; 65:2979–99.
90. Lu XG, Zhan LB, Feng BA, Qu MY, Yu LH, Xie JH. Inhibition of growth and metastasis of human gastric cancer implanted in nude mice by d-limonene. *World J Gastroenterol.* 2004; 10:2140–4.
91. Crowell PL. Prevention and therapy of cancer by dietary monoterpenes. *J Nutr.* 1999; 129:775S–8S.
92. Zhou JY, Tang FD, Mao GG, Bian RL. Effect of α -pinene on nuclear translocation of NF- κ B in THP-1 cells. *Acta Pharmacol Sin.* 2004; 25:480–4.
93. Berchtold CM, Chen KS, Miyamoto S, Gould MN. Perillyl alcohol inhibits a calcium-dependent constitutive nuclear factor - κ B pathway. *Cancer Res.* 2005; 65:8558–66.
94. Santos TG, Dognini J, Beghini IM, Rebelo RA, Verdi M, de Gasper AL, Dalmarco EM. Chemical characterisation of essential oils from *Drimys angustifolia* miers (Winteraceae) and antibacterial activity of their major compounds. *J Brazil Chem Soc.* 2013; 24(1):164–70.
95. Shahverdi AR, Rafii, F, Tavassoli F, Bagheri M, Attar F, Ghahraman A. Piperitone from *Mentha longifolia* var. *chorodictya* Rech F. reduces the nitrofurantoin resistance of strains of Enterobacteriaceae. *Phytotherapy Res.* 2004; 18:911–14.
96. Solis C, Becerra J, Flores C, Robledo J, Silva M. Antibacterial and antifungal terpenes from *Pilgerodendron uviferum* (D. Don) Florin. *J Chilean Chem Soc.* 2004; 49(2):157–61.
97. Southwell IA, Brophy JJ. Differentiation within the Australian *Tasmannia* by essential oil comparison. *Phytochem.* 1992; 31(9):3073–81.
98. Hu FB, Bronner L, Willett WC, Stampfer MJ, Bexrode KM, Albert CM, Hunter D, Manson JE. Fish and omega-3 fatty acid intake and risk of coronary heart disease in women. *JAMA.* 2002; 287(14):1815–21.
99. Ren J, Chung SH. Anti-inflammatory effect of α -linolenic acid and its mode of action through the inhibition of nitric oxide production and inducible nitric oxide synthase gene expression via NF- κ B and mitogen-activated protein kinase pathways. *J Agric Food Chem.* 2007; 55(13):5073–80.
100. Ko TF, Weng YM, Chiou RYY. Squalene content and antioxidant activity of *Terminalia catappa* leaves and seeds. *J Agricultural and Food Chemistry.* 2002; 50:5343–8.
101. Newmark HL. Squalene, olive oil and cancer risk: A review and hypothesis. *Cancer Epidem Biomar.* 1997; 6:1101–3.
102. Vetter SM, Schlievert PM. Glycerol monolaurate inhibits virulence factor production in *Bacillus anthracis*. *Antimicrob Agents Ch.* 2005; 49:1302–5.
103. Ellis CD, Lindner B, Khan A, Zahringer U, Demarco U, de Hormaeche R. The *Neisseria gonorrhoeae* 1pxLII gene encodes for a late-functioning lauroyl acyl transferase, and a null mutation within the gene has a significant effect on the induction of acute inflammatory responses. *Mol Microbiol.* 2001; 42:167–81.
104. Sun CQ, O'Connor CJ, Robertson AM. Antibacterial actions of fatty acids and monoglycerides against *Helicobacter pylori*. *FEMS Immunol Med Mic.* 2003; 36:9–17.
105. Petschow BW, Batema RP, Talbott RD, Ford LL. Impact of medium-chain monoglycerides on intestinal colonisation by *Vibrio cholerae* or enterotoxigenic *Escherichia coli*. *J Med Microbiol.* 1998; 47:383–9.
106. Ohk SH, Kuramitsu HK. A novel antibacterial agent derived from the C-terminal domain of *Streptococcus mutans* GTP-binding protein. *J Antimicrob Chemoth.* 2000; 46:95–9.
107. Epstein JB, Ransier A, Sherlock CH, Spinelli JJ, Reece D. Acyclovir prophylaxis of oral herpes virus during bone marrow transplantation. *Eur J Cancer, Part B Oral Oncology.* 1996; 32B:158–62.
108. Issacs CE, Thormar H, Pessolano T. Membrane-disruptive effect of human milk: inactivation of enveloped viruses. *J Infect Dis.* 1986; 154:966–71.
109. Isaacs CE, Thormar H. The role of milk-derived antimicrobial lipids as antiviral and antibacterial agents. *Adv Exp Med Biol.* 1991; 310:159–65.
110. Enig M. Lauric acid as antimicrobial agents: Theory of effect, scientific rationale, and dietary applications as adjunct nutritional support for HIV-infected individuals. *Nutrients and Foods in AIDS*, Watson R (Ed), CRC Press, Boca Raton.
111. Chipley JR, Story LD, Todd PT, Kabara JJ. Inhibition of *Aspergillus* growth and extracellular aflatoxin accumulation by sorbic acid and derivatives of fatty acids. *J Food Safety.* 1981; 3:109–19.
112. Bergsson G, Arnfinnsson J, Steingrímsson O, Thormar H. In vitro killing of *Candida albicans* by fatty acids and monoglycerides. *Antimicrob Agents Chemother.* 2001; 45(11):2309–3212.
113. Rayan P, Stenzel D, McDonnell PA. The effects of saturated fatty acids on *Giardia duodenalis* trophozoites in vitro. *Parasitol Res.* 2005; 97:191–200.
114. Ababouch HL, Bouqartacha F, Busta FF. Inhibition of *Bacillus cereus* spores and vegetative cells by fatty acids and glyceryl monododecanoate. *Food Microbiol.* 1994; 11:327–36.
115. Lee JY, Kim YS, Shin DH. Antimicrobial effect of linolenic acid and monoglyceride against *Bacillus cereus* and *Staphylococcus aureus*. *J Agric Food Chem.* 2002; 50:2193–9.
116. Yff BTS, Lindsey KL, Taylor MB, Erasmus DG, Jäger AK. The pharmacological screening of *Pentanisia prunelloides* and the isolation of the antibacterial compound palmitic acid. *J Ethnopharmacol.* 2002; 79:101–7.
117. Salvager JL. Surfactants types and uses. FIRP Booklet 2002; E300A: Universidad De Los Andes, Venezuela.
118. Castano G, Fernandez L, Mas R, Illnait J, Fernandez J, Mesa M, Alvarez E, Lezcay M. Comparison of the efficacy, safety and tolerability of original policosanol versus other mixtures of higher aliphatic primary alcohols in patients with type II hypercholesterolemia. *Int J Clin Pharmacol Res.* 2002; 22(2):55–6.
119. Sakulnarmrat K, Fenech M, Thomas P, Konczak I. Cytoprotective and pro-apoptotic activities of native Australian herbs polyphenolic-rich extracts. *Food Chem.* 2013; 136:9–17.
120. Sakulnarmrat K, Konczak I. Composition of native Australian herbs polyphenolic-rich fractions and in vitro inhibitory activities against key enzymes relevant to metabolic syndrome. *Food Chem.* 2012; 134:1011–19.
121. Weerakkody NS, Caffin N, Turner MS, Dykes GA. In vitro antimicrobial activity of less-utilised spice and herb extracts against selected food-borne bacteria. *Food Control.* 2010; 21:1408–14.
122. Shoeb M. Anticancer agents from medicinal plants. *Bangladesh J Pharmacol.* 2006; 1(2):35–41.