Effect of aqueous extract of *Achillea millefolium* on the development of experimental autoimmune encephalomyelitis in C57BL/6 mice

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**ABSTRACT**

Objective: *Achillea millefolium* (*A. millefolium*) is widely used as an anti-inflammatory remedy in traditional and herbal medicine. In this study, we investigated the effect of an aqueous extract from *A. millefolium* on experimental autoimmune encephalomyelitis (EAE) and on the serum cytokine levels in C57BL/6 mice.

Materials and Methods: EAE was induced in 63 C57BL/6 mice weighing 20-25 g (8 weeks old). Following immunization, the treatment protocol was initiated by using different doses of an aqueous extract from *A. millefolium* (1, 5, and 10 mg/mouse/day). Histopathologic assessments were performed by hematoxylin and eosin (H and E) and luxol fast blue (LFB) staining. Behavioral disabilities were recorded by a camera. Serum levels of interleukin (IL)-10, IL-12, and transforming growth factor (TGF)-β were measured using enzyme-linked immunosorbent assay (ELISA).

Results: On average, mice developed classical behavioral disabilities of EAE, 13.2 ± 1.9 days following immunization. Treatment of mice with *A. millefolium* led to delay the appearance of behavioral disabilities along with reduced severity of the behavioral disabilities. Treatment with *A. millefolium* prevented weight loss and increased serum levels of TGF-β in immunized mice with MOG35-55. EAE-induced mice, which were treated with *A. millefolium*, had less cerebral infiltration of inflammatory cells.

Conclusion: The results demonstrated that treatment with aqueous extract of *A. millefolium* (*A. millefolium*) may attenuate disease severity, inflammatory responses, and demyelinating lesions in EAE-induced mice. In addition, following treatment with *A. millefolium*, serum levels of TGF-β were increased in EAE-induced mice.

**KEYWORDS:** *Achillea millefolium*, cytokines, experimental autoimmune encephalomyelitis, multiple sclerosis

**Introduction**

Multiple Sclerosis (MS) is a complex disorder and despite years of focused research, the main etiological factors for pathological processes are yet to be identified.[1] Experimental autoimmune encephalomyelitis (EAE) has been used as a model for MS in animals worldwide[2] and has assisted in the identification of autoantigens associated with its pathology and progression. It is now well-documented that inflammatory responses may have a role in pathogenesis of EAE.[3] A broad range of different proinflammatory mediators including cytokines such as tumor necrosis factor (TNF)-α, interleukin (IL)-6, IL-1β, interferon (INF)-γ, and IL-17, enzymes like cyclooxygenase-2, transcription factors, nuclear factor kappa B (NF-kB) and CCAAT-enhancer binding protein-B (C/EBB) are believed to be involved in activation of cytotoxic T lymphocytes,
macrophage, and glial cells, which are responsible for the proinflammatory response in EAE. Proinflammatory responses result in destruction of oligodendrocytes.

*Achillea millefolium* (A. millefolium) or yarrow, a flowering plant from the family of Asteraceae, is widespread and frequently used as a medicinal plant worldwide. A. *millefolium* has been used as a treatment for several disorders varying from wound healing, infections diseases, pain, and gastrointestinal complaints to many other conditions. Although A. *millefolium* has been suggested as a folk remedy for the traditional treatment of central nervous system diseases, few data have been published supporting this claim. Elmann et al., reported that A. *millefolium* extract has anti-inflammatory effects on lipopolysaccharide (LPS)-activated primary cultures of brain microglial cells. Therefore, they concluded that A. *millefolium* could be beneficial in preventing or treating neurodegenerative diseases like Alzheimer and Parkinson's disease.

In another study, Molina-Hernandez et al., reported anticonflict-like actions of aqueous extract of the flowers of A. *millefolium* in rats. Also recently, Bareta et al., reported that acute and chronic oral administration of hydroalcoholic extract of A. *millefolium* exerted anxiolytic-like effects in mice.

Hence, the aim of this study was to investigate the effects of aqueous extract of A. *millefolium*, on the development of EAE in C57BL/6 mice.

**Materials and Methods**

**Animals**

Adult (8-12 weeks old) male C57BL/6 mice were purchased from the Pasteur Institute (Tehran, Iran) and were used for EAE induction. Mice were kept under optimal conditions based on the temperature, hygiene, humidity, and light (cycles of 12 h dark/light). They had free access to food and water. The method of this study was approved by local ethics committee (Ethics and Animal Care Committee of Kafsanjan University of Medical Sciences) and all experiments were conducted according to the standard ethics guidelines.

**Reagents**

Myelin oligodendrocytes glycoprotein (MOG) was purchased from Alexis Company, USA. Complete Freund’s adjuvant (CFA), cresyl fast violet, pertussis toxin, luxol fast blue (LFB), and lithium carbonate were purchased from Sigma-Aldrich, USA.

**Preparing of Plant Materials**

The plant material was collected during March 2011 from Isfahan Botany Herbarium and was identified by Dr Valiollah Mozaffarian at Botany Research Division, Research Institute of Forests and Rangelands, Tehran, Iran. A voucher specimen has been kept in Isfahan Botany Herbarium voucher specimen no. 9757. The dried aerial parts of the plant (200 g) were rinsed with distilled water, dried, ground into powder and extracted in a Soxhlet apparatus with distilled water. The solvent was evaporated under reduced pressure at 40°C. The resultant extract was then reconstituted by solving it in distilled water before being used in animals. Three of more common doses of extract were selected to use in this study (40, 200, and 400 mg/kg). Due to the fact that the most of mice weighed around 24 g, we converted the mentioned doses per mouse (1, 5, and 10 mg/mouse).

**Experimental Groups**

In this study, mice were randomly allocated into seven equal groups (seven mice per group) as follows: (1) Control group; no intervention was performed in this group, (2) EAE group: EAE was induced in this group using MOG35-55, CFA, and pertussis toxin, (3) Sham group; animals received CFA and pertussis toxin without MOG, (4-6): In these three groups, animals were initially subjected to EAE induction followed 24 h later by 1, 5, and 10 mg/mouse/day *A. millefolium* for 21 days, and 7: The healthy mice received 10 mg/mouse/day *A. millefolium* for 21 days. The last group was designed to study any effect of *A. millefolium* on overall health of the rats (see Table 1 for the treatment regime).

**Induction of EAE in Mice**

EAE was induced using a peptide corresponding to the sequence of the rodent myelin oligodendrocytes glycoprotein MOG35-55. Experimental mice received a subcutaneous injection of 250 μg MOG35-55/mouse emulsified in CFA containing 0.4 mg *Mycobacterium tuberculosis*. Animals also received 500 ng/mouse pertussis toxins immediately after immunization and 48 h later. In sham-treated group, animals received CFA and pertussis toxin, but did not receive MOG. Mice were monitored daily for weight loss and neurological signs of EAE. The severity of disease among EAE mice were scored (6 grades) based on the method reported by Onuki et al., as follows: Grade 0 = if there was no sign of disease, grade 1 = if there was partial loss of tail tonicity, grade 2 = if there was loss of tail tonicity along with tail righting disabilities, grade 3 = if there was unsteady gait and mild paralysis of one hind limb, grade 4 = if there were both hind-limb paralysis and incontinence, grade 5 = if there was quadriplegia, and grade 6 = death. The mice were humanely sacrificed 21 days after immunization with MOG35-55.

**Histological and Microscopic Assessments**

On the 21st day of EAE induction, animals were anesthetized with injection of high dose of thiopental. Blood samples were collected before intracardiac perfusion of animals with phosphate buffered saline (PBS; pH 7.4) containing 4% (w/v) paraformaldehyde solution. The whole brain was dissected out from each animal and the dissected tissues were fixed

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All EAE-induced mice received complete Freund’s adjuvant (CFA) and pertussis toxin. EAE = Experimental autoimmune encephalomyelitis.
and divided into two hemispheres. The right hemisphere was used for histological staining, whereas the left hemisphere was used for myelin staining. For histological analysis, routine histology methods were carried out to obtain morphological details of the brain tissue in EAE mice. Paraformaldehyde fixed tissues were embedded in paraffin, and serial sections (8 μm) were prepared. Sections were stained with a conventional hematoxylin and eosin (H and E) staining method. Digital images were taken using a ×40 objective. Slides were assessed in a blinded fashion for inflammation by trained experts from the Department of Anatomy, RafaSanjan University of Medical Sciences. Inflammation was scored using the following scale: 0 = No infiltrating cells were observed; 1 = few infiltrating cells were seen; 2 = numerous infiltrating cells were visible; and 3 = widespread infiltration was observed. Six serial sections of right hemisphere of each mouse in each group were scored. The processes of counting were performed in similar sections and in similar microscopic fields in all groups. Sections of the left hemisphere were incubated in 0.1% LFB dissolved in 95% ethanol and 0.05% acetic acid at 60°C for 18 h. Stained sections were differentiated in 0.05% lithium carbonate and 70% ethanol. Sections were then counterstained with cresyl fast violet, dehydrated, and mounted with entanlen. Six sections of left hemisphere of each mouse were stained with LFB and were checked for demyelinating plaque.

**Determination of Immunologic Factors**

The serum levels of IL-10, transforming growth factor (TGF)-β, and IL-12 were measured using enzyme-linked immunosorbent assay (ELISA; eBioscience, ESP). Briefly, the monoclonal antibodies against IL-10, TGF-β, and IL-12 were precoated in polyester ELISA plates and 24 h after incubation and washing were blocked in the PBS buffer containing 3% bovine serum albumin for 2 h. Samples were then added to the well plates and were incubated for 2 h. The plates were washed again and biotin-conjugated antibodies against the cytokines were added to the plates and were again incubated for 2 h. After washing the plates, horseradish peroxidase (HRP)-avidin was added and further incubated for another 30 min. Finally, the plates were washed, substrate (TMB + H₂O₂) was added and after 15 min of incubation the reaction was stopped by addition of 2N H₂SO₄. The absorption was measured using an ELISA plate reader. The sensitivity of kits was 2 pg/ml and inter- and intra-assay assessments of reliability of the kits were evaluated and produced scores of CV <14% and <0.03%, respectively.

**Data Analysis**

Data were analyzed using excel and Statistical Package for Social Sciences (SPSS). All data were reported as a mean ± standard error of mean (SEM) and a P value less than 0.05 was considered as a significant difference. Differences between the groups were measured using one-way analysis of variance (ANOVA) followed by the Dunnett’s post hoc test. Repeated measure ANOVA (RMA) was also used to compare weight changes in the days subsequent to induction of EAE.

**Results**

**Behavioral Scores**

In Group 2 (the EAE group), the first behavioral scores of EAE emerged following 13.2 ± 1.9 days of immunization. In this group, the behavioral scores increased to peak level of 3.4 ± 0.8 by 17 days following immunization (RMA, P = 0.049). Treatment with A. millefolium delayed the onset of behavioral changes compared to EAE group. In Groups 4 and 6 (received 1 and 10 mg/mouse of A. millefolium, respectively), behavioral scores of EAE were observed from 18 and 17 days after immunization, respectively. In Group 5 (received 5 mg/mouse of A. millefolium), we did not observe any behavioral score for EAE throughout the period of study [Figure 1]. The mean of behavioral score in Group 4 (P = 0.014) and Group 6 (P = 0.021) were significantly lower than Group 2 [Figure 1].

**Weight Changes**

The results showed that the mean weight of mice in Group 2 on the 21st day were significantly lower than their mean weight on the 1st day after immunization with MOG (P = 0.003). The mean weight of mice in Groups 4-6 have not changed at 21st day following immunization compared to the 1st day of immunization in each group (all P > 0.3) [Figure 2]. In Group 7 (only treated with 10 mg/mouse/day A. millefolium), we did not observe either behavioral disabilities or weight loss throughout the study, suggesting that the drug does not have a deleterious effect on general health.

**Histology**

H and E staining on the coronal sections of cerebrum in Group 2 showed massive infiltration of inflammatory cells into the brain tissue [Figure 3a]. In contrast, in groups 4-6 the infiltration of inflammatory cells into the brain tissue was decreased [Figure 3a]. The number of inflammatory cells decreased in Group 5 (P = 0.013) and 6 (P = 0.010) compared to group 2 [Figure 3b].

Groups 4-6 lesser demyelinated lesions in the brain were observed [Figure 4]. However, we did not perform statistical analysis for the number of lesion areas in the experimental groups.
Our results showed that mean serum levels of IL-10 and IL-12 were not significantly different between the experimental groups. Serum levels of TGF-β was higher in Groups 4-6, compared with Group 2, however, the difference was significant only in Group 6 ($P = 0.026$) [Table 2].

**Discussion**

The results demonstrate that treatment of EAE mice model with aqueous extract of *A. millefolium* inhibited the development of EAE symptoms and complications (both behavioral presentations and inflammation). Our results also revealed that the serum levels of IL-10 and IL-12 were not significantly altered in different experimental groups, however, the circulating TGF-β levels were significantly increased in the EAE-induced mice.
treated with 10 mg/mouse of *A. millefolium* suggesting that the effects of the extract on TGF-β may be dose-dependent, but this would need to be assessed in further research programs.

The anti-inflammatory effects of *A. millefolium* have been documented in earlier studies. **It** has been demonstrated that one of the main consequences of EAE and MS, is the infiltration of activated T cells and associated mononuclear cells such as macrophages into the central nervous system (CNS). This in turn is facilitated by a broad-spectrum of inflammatory consequences. Therefore, according to previous reports we investigated whether treatment with *A. millefolium* could attenuate the infiltration of *A. millefolium* inflammatory cells in EAE mice. Our results demonstrated that *A. millefolium* decreased the infiltration of inflammatory cells into the cerebrum of EAE treated animals. Consistent with our results, there are several reports regarding the anti-inflammatory properties of *A. millefolium*. Benedek et al., in 2007 have reported that *A. millefolium* inhibits inflammation related proteases, including human neutrophil elastase (HNE) and matrix metalloproteinases (MMPs) in vitro. In another report, Maswadeh et al., showed that a gel containing 6% *A. millefolium* extract reduced paw edema (approximately 50%) in rats. The inhibitory effects of *A. millefolium* on some mediators of inflammation such as MMPs may play an important role in the preventative effects of this extract on EAE, this is confirmed by some studies that report both MMP 2 and 9 were increased in MS. It has also been reported that oxidative stress influences the severity of MS. Furthermore, there are reports demonstrating the antioxidant properties of *A. millefolium*. Konyalioglu and Karamendere, screened 15 different species of Achillea for their antioxidant activities by using several in vitro assays on human erythrocytes and leukocytes. Therefore, it is interesting to speculate that one of the possible mechanisms by which *A. millefolium* inhibits development of EAE could occur through its antioxidant properties. However, further investigations are required to shed light on this.

With regards to the fact that TGF-β contributes to the processes of matrix synthesis, it may be reasonable to conclude that *A. millefolium* may induce the remyelination of injured neurons via upregulation of TGF-β in the EAE model. Interestingly, it has been shown that TGF-β induces the expression of chemotactic factors from microglia which are specific for the recruitment of oligodendrocyte precursor cells (OPC) in C57BL/6 mice. OPCs chemotraction to the demyelinating CNS lesions is one of the main determining factors for remyelination. Moreover, there is compelling data demonstrating that TGF-β has anti-inflammatory effects on the immune cells (such as NK cells and T cytotoxic lymphocytes). Hence, it seems that treatment with *A. millefolium* may downregulate inflammation in EAE at least partially via induction of TGF-β. On the other hand, TGF-β play dual roles when it is associated with other cytokines, therefore, future studies should be directed at looking a wider range of inflammatory cytokines (such as IL-17, interferon gamma (IFNγ), IL-23, and IL-6) to evaluate the effects of *A. millefolium* on this system.

Overall, the results showed that the serum levels of IL-12 and IL-10 were not changed among the studied groups. It has been shown that IL-17A has antagonistic effect on the IL-12 and IL-10 secreting cells. Moreover, recent studies demonstrated that IL-17A is one of the main cytokines involved in the induction of the EAE, thus, it is difficult to reconcile the reported influence of IL-17A on EAE with our data that showed no changes in the downstream cytokines, IL-12 and IL-10. Further studies need to clarify possible interaction between these cytokines in EAE-induced mice.

To conclude *A. millefolium* can inhibit the development of EAE in mice. Future clinical studies on the effect of *A. millefolium* in patients with MS may be planned based on the findings of this study.

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**References**
